

PROTOLYTIC REACTIONS AND THE FUNCTIONING OF F-ATPases

F-ATPases synthesize ATP driven by a protonmotive, in some cases by a sodiummotive force. Their construction is bipartite with a membrane integral channel portion and a peripheral catalytic portion, both consisting of more than 20 polypeptides. These are remarkably conserved in tertiary and quaternary but not in primary structure. Exchange of subunits between remote organisms as *E. coli*, *Synechocystis* sp. FCC 6803 and spinach by proteinchemical or genetical manipulations has led to partially or fully functional chimeric enzymes. It is conceivable that they function as electrochemical-to-mechanical-to-chemical, or proton-to-conformation-to-ATP transducers. The extreme proton selectivity of the channel portion of the chloroplast enzyme ($H^+/K^+ > 10^6$) sharply contrasts with the sodium dependence of *P. modestum*. This observation also points to a non-chemical, indirect action of protons (1-5). We have attempted to resolve partial reactions of the proton by application of flash spectrophotometry to the chloroplast enzyme in the thylakoid membrane, using both pH and voltage probes. A clue came from studies on a proton slip in the absence of nucleotides. F-ATPases contain a total of six nucleotide binding sites that can be grouped into two subsets, catalytic and non-catalytic ones, according to their specificities and affinities. Increasing the concentration of ADP the enzyme is transformed between three states with different proton conductance: uncoupled ("proton slip") \Rightarrow closed \Rightarrow coupled (ATP-synthesis). Proton slip is inhibited by the binding of ADP ($K=200nM$, plus P) to a first binding site. Proton flow coupled to ATP-synthesis is initiated by the binding of another molecule of ADP ($K=7\mu M$, plus P) to at least another site. Already the first site belongs into the subset of potentially catalytic sites. We interpret the above state transition as a consequence of an alternating site mechanism of catalysis. A mechanical model is presented which links proton flow to the extrusion of spontaneously synthesized and bound ATP. This model explains ATP synthesis under multisite and unisite conditions as well as ATP hydrolysis driven proton pumping. Under conditions of proton slip we have demonstrated for the first time that the binding, the transmembrane transport, and the extrusion of protons can be selectively affected by nucleotides and inhibitors of the enzyme. Evidence for a "proton well" construct (Mitchell) at the inlet and the outlet side of the enzyme is presented (2, 6, 7).

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