

Photosynthetic Water Oxidation at High O₂ Backpressure Monitored by Delayed Chlorophyll Fluorescence[†]

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ABSTRACT: The atmospheric dioxygen is produced by photosynthetic organisms. This light-driven process culminates in what appears as one step: a four-electron abstraction from two water molecules bound to the Mn₄Ca complex of photosystem II. Recently, an intermediate of the O₂-producing reaction sequence was stabilized by elevated oxygen backpressure and detected by UV flash photometry [Clausen, J., and Junge, W. (2004) *Nature* 430, 480]. We scrutinized its properties by delayed chlorophyll fluorescence measurements. Half-suppression of oxygen evolution was observed at a similar O₂ pressure of 2.3 bar, as previously, now with photosystem II membrane particles from spinach, without artificial electron acceptors, and at a high signal-to-noise ratio. The data are tentatively interpreted as the stabilization of a 2-fold oxidized state of the catalytic center (S₂^{*}) with bound peroxide and its slow conversion into the normal S₂ state by the release of peroxide.

Plants, algae, and cyanobacteria oxidize water and produce dioxygen in photosystem II (PSII).¹ The catalytic center comprises a Mn₄Ca complex bound to the D1 subunit of PSII, the atomic structure of which is now emerging from crystallographic (1, 2) and X-ray absorption spectroscopy studies [(3, 4) and references therein]. The growing structural information calls for in-depth investigations on the crucial reaction sequence of oxygen evolution to unravel its still enigmatic chemical mechanism.

Clocked forward by the absorption of four quanta of light, the catalytic center acquires increasingly higher oxidation states (5) (starting from S₀ to reach S₁ (dark stable), S₂, S₃, and S₃Y_Z^{ox}, wherein Y_Z denotes a special, redox-active tyrosine). The terminal dark reaction (S₃Y_Z^{ox} → S₀) closes the cycle and is coupled to the release of dioxygen. One view is that, upon reaching the highest oxidation state, four electrons are abstracted from two molecules of bound water to liberate dioxygen so that S₃Y_Z^{ox} is reduced to S₀. Until recently, the latter reaction appeared as a single step without a detectable intermediate. Evidence for the existence of such an intermediate has been provided in a study employing

elevated oxygen pressure (6). An O₂ pressure of only 2.3 bar (about 10-fold higher than ambient) half-inhibited the liberation of oxygen by stabilizing an intermediate. This intermediate was tentatively ascribed to a 2-fold-oxidized manganese complex (similar to state S₂), which possibly carries a bound peroxide.

In this investigation, we used recombination chlorophyll fluorescence after laser-pulse excitation (the so-called delayed fluorescence, DF) to monitor the redox state of the catalytic center at elevated oxygen pressure. DF stems from the reversal, in a thermally activated process, of the flash-light-induced charge separation in PSII (7). Its intensity reflects the free energy of the reached charge-separated state with respect to the excited antenna state (7). The various kinetic phases of DF transients reflect the partial reactions of the electron-transfer chain Mn → Y_Z → P₆₈₀⁺, wherein Mn denotes the catalytic metal complex and its ligands, Y_Z denotes the Tyr-161 on the D1 subunit, and P₆₈₀⁺ denotes the photooxidized chl_a moiety that drives these reactions (for a comprehensive description of the DF method as applied to PSII see ref 7). The major advantage of DF over UV (see ref 6) and X-ray flash spectroscopy (see ref 4) is its very high signal-to-noise ratio. It allows single-shot experiments at microsecond time resolution even with scattering samples, like PSII membrane particles from spinach, as used in this study.

By the DF technique, we corroborated the previously found stabilization of an intermediate by elevated oxygen pressure (6) but now with PSII of a higher plant in its native membrane and without the complication of added electron acceptors. A plausible assumption is that this intermediate is equivalent to (H₂O₂)S₂^{*}Y_Z. Presumably, this state slowly decays, within less than 700 ms, perhaps to yield the normal 2-fold oxidized S₂ state of the catalytic center by releasing the bound peroxide. This finding together with kinetic evidence for another intermediate, as detected by time-

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¹ Abbreviations: chl, chlorophyll; DF, delayed-fluorescence emitted by the chlorophylls of PSII; P₆₈₀, primary electron donor of PSII; PSII, photosystem II, S_n, oxidation states in the S-state cycle of water oxidation; Y_Z, tyrosine-161 of the D1 protein of PSII.

resolved X-ray absorption spectroscopy (8), might offer a clue to unravel the whole reaction sequence of oxygen evolution.

MATERIALS AND METHODS

PSII-enriched membrane particles were prepared from market spinach with 1 M glycine betaine present as a stabilizer in all media (9) and frozen at -80°C until use. Their O_2 -evolution activity determined by a Clark electrode was $\sim 1200 \mu\text{mol}$ of O_2 (mg of chl) $^{-1} \text{h}^{-1}$. For DF measurements, PSII membranes were thawed and diluted at $20 \mu\text{g}$ of chl mL^{-1} in buffer B (1 M glycine betaine, 10 mM NaCl, 5 mM CaCl_2 , and 25 mM MES at pH 6.5). The presence of about 5 plastoquinone molecules per PSII allowed ~ 10 turnovers/PSII reaction center. Thus, the addition of an artificial electron acceptor was not necessary.

Kinetic delayed chlorophyll fluorescence measurements (DF) were performed at room temperature ($\sim 20^{\circ}\text{C}$) with the setup described in ref 7 using a gated photomultiplier for monitoring DF transients. The same optical cell with fused quartz windows sustaining pressures up to 30 bar as described in ref 6 was used for the DF experiments. Dark-adapted PSII samples were excited by trains of saturating flashes from a Continuum Minilite-II laser (frequency-doubled, Q-switched Nd:YAG, $\lambda = 532 \text{ nm}$, full width at half-maximum of $\sim 5 \text{ ns}$, flash spacing of 700 ms, attenuated to 2 mJ cm^{-2}). The laser beam was shaped by appropriate optics to homogeneously illuminate the whole sample volume. DF transients were corrected for a small DF contribution of the quartz windows as outlined in ref 7. Transients were recorded on a personal computer equipped with a 20 MHz A/D card (Adlink) and in-house software using an averaging algorithm resulting in logarithmic spacing of data points starting at $20 \mu\text{s}$ after the laser flash (electrical bandwidth set by a Tektronix amplifier to 1 MHz).

PSII samples were equilibrated with elevated oxygen (or He) pressure as follows: (1) 100 mL of buffer B was pre-equilibrated at the desired $p(\text{O}_2)$ for 30 min in a stainless-steel pressure cell under stirring. (2) 3 mL of the pre-equilibrated buffer was filled in the cuvette (optical path length of 1 cm) containing a magnetic stirring bar; the cuvette then was mounted into the optical cell. (3) The cell was pressurized and further equilibrated for 10 min under stirring. (4) An aliquot of the PSII membranes ($\sim 50 \mu\text{L}$) was injected by a syringe into the pressurized cell, and the solution was stirred in the cuvette for 20 min in darkness before application of the laser flashes. O_2 gas was of the highest available purity.

RESULTS

Figure 1 shows DF transients for up to 10 ms after the 1st, 2nd, and 3rd Laser flash, which was applied to dark-adapted PSII membrane particles at ambient (i.e., 0.21 bar) (A) and 20 bar (B) partial pressure of oxygen. The initial extent of DF (at $20 \mu\text{s}$) after flashes 1 and 2 was unchanged by elevated oxygen pressure, and most of the decay occurred rapidly, within microseconds (Table 1). We conclude that the electron-transfer $\text{Mn}_4 \rightarrow \text{Y}_Z \rightarrow \text{P}_{680}^+$ during the $\text{S}_1 \rightarrow \text{S}_2$ and $\text{S}_2 \rightarrow \text{S}_3$ transitions is not strongly affected by elevated $p(\text{O}_2)$.

The DF transient on the 3rd flash depends strongly on $p(\text{O}_2)$. At ambient $p(\text{O}_2)$ of 0.2 bar, the rapid initial decay

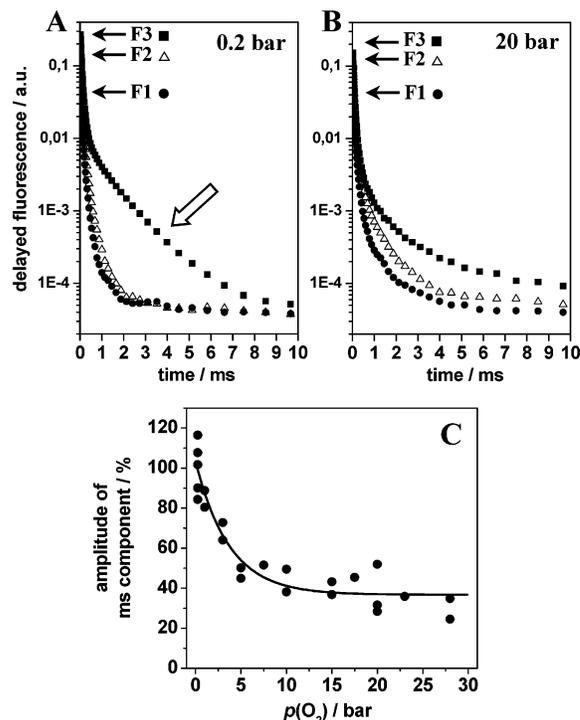


FIGURE 1: Delayed fluorescence transients recorded with PSII membranes on laser flashes 1, 2, and 3 at ambient $p(\text{O}_2)$ of (A) 0.2 bar and (B) 20 bar. The arrow in A emphasizes the DF decay with $\tau \approx 1.2 \text{ ms}$ attributable to the O_2 -evolving transition on flash 3. Each transient represents the average of three measurements. Note the logarithmic scale of the y axis. (C) Normalized intensities integrated from 0.5 to 5 ms of the DF transients on flash 3 as a function of $p(\text{O}_2)$. The line represents a single-exponential simulation with a $p(\text{O}_2)_{1/2}$ of 2.3 bar.

Table 1: Amplitudes (I_i) and Relaxation Times (τ_i) of a Triexponential Fit of the Decay of the DF Transients Shown in Figure 1^a

flash number	$p(\text{O}_2)/\text{bar}$	I_1	τ_1	I_2	τ_2	I_3	τ_3
1	0.2	19.27	0.071	1.47	0.25	0.03	4.3
	20	19.63	0.072	0.81	0.39	0.04	4.4
2	0.2	62.03	0.053	4.21	0.25	0.03	4.1
	20	53.11	0.064	1.68	0.53	0.05	4.6
3	0.2	95.93	0.074	3.95	1.17	0.12	4.5
	20	69.39	0.067	2.01	0.59	0.37	4.7

^a The amplitudes are given in percent of the total summed amplitudes for the 3rd flash transient at 0.2 bar O_2 ; relaxation times are given in milliseconds. Oxygen pressure and flash number are as indicated.

of the DF occurred with a relaxation time of $\sim 70 \mu\text{s}$ and was completed after less than $500 \mu\text{s}$. This DF decay in part reflects the slowest component of the electron transfer from Y_Z to P_{680}^+ , which presumably is rate-limited by relaxation processes at the level of Y_Z (10). It was followed by a prominent slower decay phase (Figure 1A, arrow) with a relaxation time τ_2 of $\sim 1.2 \text{ ms}$ (Table 1). Because this phase was observed on the 3rd flash and its magnitude oscillated with a period of four (see below), it is attributed, as previously indicated (7), to the reduction of $\text{S}_3\text{Y}_Z^{\text{ox}}$ during the O_2 -evolving reaction.

At 20 bar O_2 , on the 3rd flash, the decay of the largest DF portion ($\tau \approx 70 \mu\text{s}$) was similar to that at 0.2 bar (Table 1). The fact that the rapid decay was not affected by elevated oxygen pressure is in line with the previously reported reduction of P_{680}^+ in nanoseconds at 20 bar oxygen pressure as recorded by absorption transients at 827 nm (11, 12). High

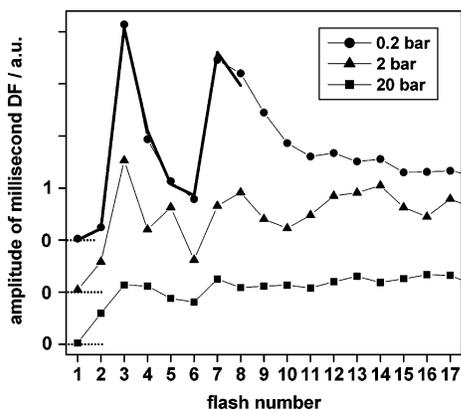


FIGURE 2: Flash patterns of the DF intensities integrated from 0.5 to 5 ms, at 0.2, 2.0, and 20 bar O₂. The integrated intensities after flash 1 were set to 0. The thick line was calculated with a miss factor of 7%, an initial population of 20% S₀ and 80% S₁, and a small exponentially rising offset (not shown) possibly because of the reduction of the quinone pool as function of the flash number.

$p(\text{O}_2)$ affected, however, slower DF phases, decaying in the time range between 0.5 and 5 ms (Figure 1C). The amplitude of these DF phases was diminished to about 35%. The decay time was accelerated from $\tau_2 \approx 1.2$ ms at ambient pressure to ~ 0.55 ms at 20 bar O₂ (Table 1). The DF transients on flash 4 and on higher flash numbers at 20 bar O₂ were very similar to that on flash 3 (data not shown). DF transients measured at 20 bar He (not shown) were essentially identical to those at 0.2 bar O₂, confirming that hydrostatic pressure per se does not affect the Mn \rightarrow Y_Z^{ox} electron transfer, as previously observed (6). At 20 bar O₂, also on the 2nd flash, a minor portion of the DF decayed with a similar τ_2 of about 0.55 ms (Table 1) as observed on flash 3 (see also the next section).

The integrated DF amplitudes measured between 0.5 and 5 ms after each flash of a series and at three values of $p(\text{O}_2)$ are plotted in Figure 2. The pronounced period-of-four oscillation observed at 0.2 bar is well-described by a “miss” factor of 7% and assuming an initial population of 20% S₀ and 80% S₁ (Figure 2). Such parameters are frequently observed in flash experiments using highly active PSII membrane particles (13). After the 10th flash, the oscillations ceased because of complete reduction of the 5 plastoquinone molecules/PSII (14).

Already at 2 bar O₂, the extent of the DF in the millisecond time range was half-diminished, although still oscillatory (Figure 2). There were further differences of the flash pattern. The millisecond decay component was not only observed on flash 3 but also on flashes 2 and 5. Possibly, a fraction of centers underwent the O₂-evolving transition on these flashes (see the Discussion). Also noteworthy, at 2 bar O₂, oscillations were still detectable at flash numbers exceeding 10. This we attribute to the incomplete turnover of centers at lower flash numbers, suggesting that, on the average, less than one electron was stably transferred to the quinones.

At 20 bar O₂, the period-of-four oscillation was absent (Figure 2). Still, a small essentially nonoscillating DF amplitude in the 0.5–5 ms time range was detectable from the 3rd flash onward, suggesting relatively slow reduction of Y_Z^{ox} (slower than in the S₂ \rightarrow S₃ transition induced by the 2nd flash). Its rate was similar on the 3rd and all later flashes (not documented), suggesting that the same reactions are in-

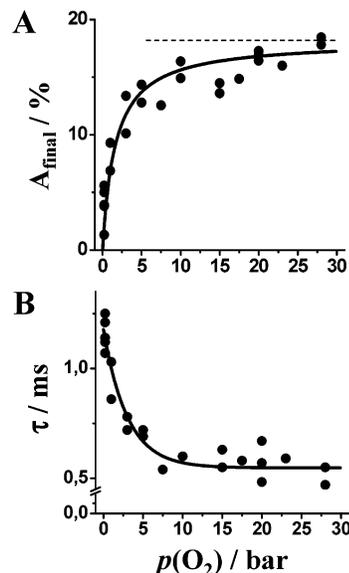


FIGURE 3: (A) Fraction of A (S₃Y_Z^{ox}) remaining in equilibrium after electron transfer in the O₂-evolving transition calculated according to $A_{\text{final}} = I_3/(I_2 + I_3)$. DF amplitudes (I_i) were obtained by triexponential simulations (see Table 1). The sum ($I_2 + I_3$) corresponds to the initial DF amplitude in the presence of S₃Y_Z^{ox}. I_2 denotes the magnitude of the millisecond DF kinetic component; I_3 is the level reached after S₃Y_Z^{ox} reduction, i.e., essentially after S₀ formation at 0.2 bar O₂ and after S₂* formation at 20 bar O₂. The smooth line represents a fit according to eq 2, with parameters given in the Discussion. The dashed line denotes the equilibrium fraction of A at infinite $p(\text{O}_2)$. (B) Apparent time constant, τ , of the increase of A. The line represents an exponential simulation with $p(\text{O}_2)_{1/2} = 2.3$ bar.

duced by these flashes. The comparatively high DF amplitude at 10 ms observed from the 3rd flash onward indicates increased recombination losses. Therefore, we suggest that the state formed in the millisecond time range after the 3rd and later laser flashes at 20 bar O₂ decayed by recombination toward the S₃ state in the 700 ms interval between flashes so that from the 3rd flash onward each flash initiated the same sequence of events (and thus the same DF decay).

The DF decay was simulated by a sum of three exponentials (see Table 1) for DF transients induced by the 3rd flash and measured at $p(\text{O}_2)$ values ranging from 0.2 to 28 bar. The simulation results facilitated calculation of the equilibrium fraction of the initial state A (here assumed to be S₃Y_Z^{ox}) formed after the 3rd flash (Figure 3A, A_{final} calculated as detailed in the caption, see also the Discussion). A_{final} was <5% at 0.2 bar O₂ but increased to close to 20% at saturating $p(\text{O}_2)$. Concomitantly, the apparent relaxation time τ_2 (see Table 1) of the decay of A decreased from ~ 1.2 to ~ 0.55 ms (Figure 3B) (A slower DF relaxation time τ_3 of ~ 4.5 ms was present on all flashes. This phase mostly reflects reoxidation of Q_A⁻, which was found to be relatively slow in the used PSII membrane particles (15) and appears to be pressure-independent). A more complex simulation approach of the DF transients on flash 3, e.g., using four exponentials, yielded a less pronounced decrease of τ_2 and a higher equilibrium fraction A_{final} . Employing a constant relaxation time τ_2 of 1.2 ms in the simulations yielded a maximum of A_{final} close to 40% (not documented).

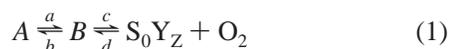
DISCUSSION

UV spectroscopy on isolated PSII core particles from cyanobacteria subjected to elevated oxygen pressure has

provided evidence for at least one intermediate in the O₂-forming transition that tentatively has been ascribed to (H₂O₂)S₂*Y_Z, the 2-fold oxidized metal cluster with a bound peroxide (6). In this study, we applied an alternative method, delayed chlorophyll fluorescence, to corroborate and extend the above finding. Delayed fluorescence is distinguished by its high signal-to-noise ratio and applicability also to native PSII membranes (7). We find the pressure-dependent changes in the 3rd-flash DF transients of PSII membrane particles from plant material (spinach) to be compatible with the presence of an intermediate in the oxygen-evolving step, and we have used the same consecutive reaction scheme as in ref 6 for their interpretation. The common result of both approaches, UV spectroscopy and DF, is 50% inhibition of oxygen evolution already at a moderate *p*(O₂) of 2.3 bar.

On the Origin of the Millisecond Phase on Flashes 2 and 5 at Elevated p(O₂). The observation of a DF phase with a millisecond decay time on the 2nd and 5th flash at elevated *p*(O₂) is explainable by assuming that a S₂* state is formed that converts to the normal S₂ state. One option for the S₂* → S₂ conversion is the release of a peroxide bound to the metal complex in S₂*. This S₂ formation may occur via two alternative pathways. (i) Centers initially in the S₀ state are converted via S₂* → S₂ in the dark. Assuming that about 20% of PSII is in S₀ in dark-adapted samples, the millisecond phase on the 2nd flash can be explained by this pathway. (ii) Centers present in state S₂* after the 3rd flash are converted to S₂ in the time interval of 700 ms between flashes 3 and 4. The millisecond phase on the 5th flash may be explainable by either pathway. We feel that the present DF results represent a first indication that O₂ exerts a true backpressure also on the fully relaxed S₀ state, driving it backward via S₂* to yield S₂. Proving the suggested S₂* formation and peroxide release remains an important challenge for future research.

Energetics of the Final Transition during Water Oxidation. The DF component with τ₂ ≈ 1.2 ms at ambient *p*(O₂) reflects the decay of the initial state in the below reaction scheme (denoted as A). It is assigned to the S₃Y_Z^{ox} state, which decays in the O₂-forming transition in parallel to manganese reduction. At high *p*(O₂), we find this decay to be accelerated (τ ≈ 0.55 ms), in line with the acceleration of the millisecond phase in the UV experiments (6, 11, 12). As in ref 6, the following two-step reaction scheme is employed:



where the intermediate state, B, is tentatively identified with S₂*Y_Z, a 2-fold reduced state of the catalytic metal center and *a*, *b*, *c*, and *d* are the rate constants of the respective forward and reverse reactions. The *p*(O₂) dependence of A_{final}, the fraction of A remaining in equilibrium, then is given by

$$A(p(O_2))_{final} = \left[1 + \frac{a}{b} \left(1 + \frac{c}{d \cdot p(O_2)} \right) \right]^{-1} \quad (2)$$

Simulation of the data points in Figure 3A based on eq 2 yielded ratios of *a*/*b* = 5.0 and *c*/*d* = 1.2 bar (−). Accordingly, with Δ*G*^{o'} = −*k*_B*T* ln *K* (*k*_B = 0.086 meV K^{−1}, *T* = 293 K), a free-energy change of Δ*G*^{o'}(A → S₂*Y_Z) = −40 meV (for *K* = *a*/*b*) is calculated. This value represents

the lower limit; an upper limit of −10 meV is obtained for an invariable time constant of 1.2 ms in the DF simulations. The respective value of +17 meV reported in ref 6 may be the upper limit; incorporation of the miss factor in the UV data analysis may render Δ*G*^o negative. In summary, S₂* formation is likely a downhill process, so that this state can accumulate to significant amounts.

For Δ*G*^{o'}(S₂*Y_Z → S₀Y_Z + O₂), values of −45/+71 meV at 0.2/20 bar O₂ are calculated [for *K* = *c*/*d**p*(O₂)]; the magnitude of ΔΔ*G*^{o'} (the Δ*G* difference for 20 bar − 0.2 bar) of ≥100 mV is only weakly affected by the used simulation approach. In line with the conclusions reached in ref 6, oxygen evolution at 20 bar O₂ is an energetically unfavorable uphill process, meaning that it practically does not proceed any more.

The above estimates by DF for the Gibbs free-energy profile of the reaction sequence (see eq 1) in part deviate from previous figures based on UV data (ref 6). The differences may or may not result from a limited signal-to-noise ratio (in particular in the UV measurements) or from different material used in the UV (cyanobacterial PSII-core complexes) and in the DF studies (plant PSII membranes). More important than these differences is the common result, namely, the pronounced lowering of the driving force for the oxygen-evolving reaction by “O₂ backpressure” as evident from half-inhibition at only 10-fold higher oxygen pressure than ambient.

In conclusion, by exerting an oxygen backpressure affecting specifically the O₂-evolving step, we have a tool at hand to stabilize an intermediate of the water-splitting reaction for further spectroscopic analysis. Recent time-resolved X-ray absorption experiments provide evidence for another short-lived intermediate of the Mn complex, which is observable even at ambient oxygen pressure (8). The step-by-step dissection of the “inner sanctum” of PSII, the O₂-producing reaction, for its in-depth mechanistic understanding now may be within reach.

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