cambridge.org/qrb

Review

Cite this article: Junge W (2019). Oxygenic photosynthesis: history, status and perspective. *Quarterly Reviews of Biophysics* https://doi.org/10.1017/S0033583518000112

Received: 30 November 2018 Revised: 8 December 2018 Accepted: 10 December 2018

Keywords:

Photosynthesis; water oxidation; oxygen; manganese cluster

Author for correspondence: Wolfgang Junge, E-mail: junge@uos.de

Oxygenic photosynthesis: history, status and perspective

Wolfgang Junge

Universität Osnabrück, 49069 Osnabrück, Germany

Abstract

Cyanobacteria and plants carry out oxygenic photosynthesis. They use water to generate the atmospheric oxygen we breathe and carbon dioxide to produce the biomass serving as food, feed, fibre and fuel. This paper scans the emergence of structural and mechanistic understanding of oxygen evolution over the past 50 years. It reviews speculative concepts and the stepped insight provided by novel experimental and theoretical techniques. Driven by sunlight photosystem II oxidizes the catalyst of water oxidation, a hetero-metallic $Mn_4CaO_5(H_2O)_4$ cluster. Mn_3Ca are arranged in cubanoid and one Mn dangles out. By accumulation of four oxidizing equivalents before initiating dioxygen formation it matches the four-electron chemistry from water to dioxygen to the one-electron chemistry of the photo-sensitizer. Potentially harmful intermediates are thereby occluded in space and time. Kinetic signatures of the catalytic cluster and its partners in the photo-reaction centre have been resolved, in the frequency domain ranging from acoustic waves via infra-red to Xray radiation, and in the time domain from nano- to milli-seconds. X-ray structures to a resolution of 1.9 Å are available. Even time resolved X-ray structures have been obtained by clocking the reaction cycle by flashes of light and diffraction with femtosecond X-ray pulses. The terminal reaction cascade from two molecules of water to dioxygen involves the transfer of four electrons, two protons, one dioxygen and one water. A rigorous mechanistic analysis is challenging because of the kinetic enslaving at millisecond duration of six partial reactions (4e⁻, 1H⁺, 1O₂). For the time being a peroxide-intermediate in the reaction cascade to dioxygen has been in focus, both experimentally and by quantum chemistry. Homo sapiens has relied on burning the products of oxygenic photosynthesis, recent and fossil. Mankind's total energy consumption amounts to almost one-fourth of the global photosynthetic productivity. If the average power consumption equalled one of those nations with the highest consumption per capita it was four times greater and matched the total productivity. It is obvious that biomass should be harvested for food, feed, fibre and platform chemicals rather than for fuel.

Introduction

Cyanobacteria, algae and plants carry out oxygenic photosynthesis. They produce oxygen (O_2) from water and take up carbon dioxide (CO_2) to yield biomass. Cell respiration reverses this process. Oxygenic photosynthesis has formed the oxygen we breathe and most of the biomass we use as food, feed, fibre and fuel. Solar driven oxygen evolution started very early in evolution. A dramatic rise of the atmospheric oxygen level (by more than four orders of magnitude), the Great Oxygenation Event (GOE), dates 2.4 billion years back from now (Bekker *et al.*, 2004; Kump, 2008). Both geological (Planavsky *et al.*, 2014) and genomic evidence (Cardona *et al.*, 2015) have suggested that ancestors of cyanobacteria might have started oxygenic photosynthesis half a billion years before the GOE. The stabilization of land masses and the emergence of land plants about half a billion years back from now caused another rise of the atmospheric oxygen content. Photosynthetic produced biomass and oxygen have powered the vast emergence of animal life. The time window of *Homo sapiens* is just a blip on these time scales. Its impact on the consumption of photosynthesis products is tremendous, as will be discussed at the end of this paper.

Several authors have comprehensively reviewed the respective momentary status of knowledge on the mechanism of oxygenic photosynthesis. They are cited in the text. The present paper scans how our knowledge on this fundamental process has unfolded over the past 50 years. Freshmen may start by reading the excellent book on photosynthesis by Bob Blankenship (Blankenship, 2014).

Figure 1 illustrates the present concept of the key elements of the primary processes of photosynthesis in green plants (photosystem II (PSII), PSI, $cytb_6f$ and the adenosine triphosphate (ATP) synthase). It is meant as a scaffold for the reader when travelling through the originally foggy, and then increasingly detailed and complicated terrain that follows.

CAMBRIDGE UNIVERSITY PRESS

Fig. 1 - Colour online



Fig. 1. Key proteins of the primary processes of photosynthesis in green plants (PSII, PSI, $cytb_6f$ and the ATP synthase), electron transfer (red), proton transfer (purple) and products (O₂, NADPH and ATP) (Junge and Nelson, 2015).

Pioneering studies

Photosynthetic oxygen production from water

In 1772 Joseph Priestley reported a first systematic study on gas turnover between plants and animals (Priestley, 1772). A mouse confined in a sealed vessel suffocated if not backed up by a plant. It has meant that the mouse produces 'bad air' and the plant restores 'vital air'. In modern terms, plants take up carbon dioxide (CO₂) and produce oxygen (O₂), while animals inhale O2 and exhale CO2. Hundred years after Priestley Theodor Engelmann (Engelmann, 1881) introduced a less harmful bioassay for oxygen, namely the flocking behaviour of oxygen-loving, motile bacteria around illuminated chloroplasts. It yielded the first action spectrum of oxygenic photosynthesis (with typical chlorophyll peaks). Half a century later the great biochemist Otto Warburg (Nobel Prize 1931) studied by manometry the turnover of O₂ and CO₂ in respiring cells and tissues (Warburg, 1922). Robert Emerson and William Arnold used his technique to detect O2-production by illuminated algae (Emerson and Arnold, 1932). They pioneered excitation of photosynthesis by short flashes of light (duration some 10 µs). Varying both the repetition rate of flashes and their energy they obtained two seminal results (Emerson and Arnold, 1932). (i) The rate of oxygen evolution is limited by a temperature-sensitive (dark) step of some 10 ms duration, and (ii) the maximum production per flash is 1 mol O₂ per 2500 mol chlorophyll. The relevance of these findings could only later be fully appreciated (see below).

The minimal balanced equation of oxygenic photosynthesis is $2H_2O + CO_2 = O_2 + (CH_2O)$. The substrate for oxygen production, water or carbon dioxide, was under debate. Robin Hill studied photosynthetic oxygen production in freshly isolated chloroplasts by another bio-assay, the spectral shift of added oxy-/deoxy-haemoglobin. Upon ageing of chloroplasts both, oxygen production and carbon dioxide consumption faded out. The addition of ferric ions as oxidant restored oxygen production but not carbon dioxide uptake. Hill took the decoupling of the former from the latter as evidence that plants 'do not evolve oxygen from carbon dioxide' (Hill, 1939). Using heavy oxygen as a tracer Martin Kamen and collaborators found the 18O/16O-ratio of the evolved oxygen identical with that of the water'. The obvious conclusion has been that O₂ originates solely from water (Ruben et al., 1941) (for a modern remake see Clausen et al. (2005a)). Otto Warburg had meanwhile perfected the manometric monitoring the exchange of CO₂ and O₂ in photosynthesis and respiration (Burk et al., 1951). Having observed a stimulating role of bicarbonate on photosynthetic oxygen production he claimed that oxygen results from bicarbonate and not from water (Warburg et al., 1965). The origin of the stimulating effect of

77

78

79

80

81

82

83

84

85

86

87

88

89

90

91

92

93

94

95

96

97

98

66

 CO_2 on PSII that drove Warburg to this erroneous view, has only been settled much later (reviewed by Govindjee and his colleagues (Shevela *et al.*, 2012)) and the regulatory function of CO_2 was detailed (Koroidov *et al.*, 2014; Brinkert *et al.*, 2016).

Electron transport, oxygen production, proton transport and ATP synthesis

Lou Duysens (Duysens et al., 1956), Bessel Kok (Kok, 1956) and Horst Witt (Witt and Moraw, 1959) started to characterize a wealth of spectroscopic signatures of photosynthetic electron transfer. Excitation with short flashes allowed resolving partial reactions in time (Kok, 1956; Witt et al., 1961a). Two PSs operate in series (Duysens et al., 1961; Witt et al., 1961b). PSII produces oxygen and delivers electrons to PSI, which in turn provides electrons for the reduction of CO₂ (see Fig. 1). The rate limiting step of the whole electron transport chain is located between the two PSs. Its duration (about 10 ms) is compatible with the one found for oxygen production by Emerson and Arnold (Emerson and Arnold, 1932). The sensitizers of both PSs are chlorophyll-a constructs, which have been coined after the wavelength of their respective red absorption peak. P700 (Kok and Gott, 1960) drives PSI and P680 (Döring et al., 1967) drives PSII, respectively. These biophysical data were then not readily appreciated by some biochemists. When Warburg was confronted with Witt's detailed reaction scheme in 1962, he mused: 'Could you tell us how the chemical mechanism of photosynthesis can be described on the basis of your spectroscopic observations?' Witt countered his eminent critic, the pioneer of oxygen detection, by observing that 'it would be difficult to deduce the mechanism of a combustion engine based only on sniffing the exhaust' (see Junge and Rutherford (2007)). From their then limited perspective both were of course right (see below).

At that time it had been accepted that the chlorophylls and 99 therewith the primary photosynthetic activity is embedded in the 100 tightly folded inner membranes of chloroplasts, forming thylakoids 101 (Menke, 1962). Peter Mitchell (Nobel Prize 1978) hypothesized a 102 new role of this inner membrane system of chloroplasts and like-103 wise the cristae membrane of mitochondria (Mitchell, 1961, 104 1966). He envisaged that the membrane is proton tight such that 105 zigzagging electron/hydrogen transport generates a pH-difference 106 (Mitchell, 1961), and transmembrane voltage (Mitchell, 1966), in 107 sum coined as protonmotive force. The backflow of protons is con-108 fined to the ATP synthase where it drives ATP-synthesis. Essentials 109 of Mitchell's hypothesis were soon experimentally established for 110 the thylakoid membrane. André Jagendorf subjected broken chlo-111 roplasts to an acid/base-jump (pH-jump) and obtained ATP 112 (Jagendorf and Uribe, 1966). The author with Horst Witt discov-113 ered the electrochromic origin of certain absorption transients of 114 intrinsic chloroplast pigments (Junge and Witt, 1968). Upon exci-115 tation with flashing light they observed very rapid voltage genera-116 tion by both PSs and slower proton pumping (Schliephake et al., 117 1968). Subsequent studies of the author with Bernd Rumberg 118 have revealed that the flash generated voltage decays in synchrony 119 with ATP synthesis (Junge, 1970; Junge et al., 1970). Titration of 120 the voltage decay with the extremely powerful ionophore gramici-121 din revealed that the electrified membrane contains at least 10⁵ 122 chlorophyll molecules (Junge and Witt, 1968), i.e. more than 100 123 electron transport chains. This number was later raised up to 124 more than 10^7 chlorophyll molecules (Schönknecht *et al.*, 1990). 125 In other words, the intricately folded thylakoid membrane within 126 a chloroplast forms one contiguous, i.e. simply connected sheet 127

(Junge, 1977; Schönknecht *et al.*, 1990). It implies that almost all copies of the electron transport chain contribute driving force to all copies of ATP synthase. The lateral separation of PSII and PSI (and ATP synthase) in different areas of the folded thylakoid membrane (Boardman and Anderson, 1964) apparently does not impair their ability for concerted electron and proton transport. Losses of the chemical portion of protonmotive force between laterally separated proton pumps and ATP synthase were only later discovered. They are minor both in mitochondria (Rieger *et al.*, 2014) and in chloroplasts (Sjoholm *et al.*, 2017). Top efficiency of ATP synthesis has apparently been sacrificed for the sake of tight packing of the coupling membrane, both in chloroplasts and mitochondria.

Photosystem II with the oxygen evolving complex (BC)

Four-electron chemistry of water oxidation linked to one-electron photo-physics

In 1968 Anne and Pierre Joliot have lifted studies of photosynthetic oxygen evolution to a kinetic level by introducing a rapidly responding oxygen electrode (Joliot and Joliot, 1968). When they excited dark adapted Chlorella cells with a series of short light flashes oxygen production peaked at flash number 3 and continued with damped oscillations of period four (Joliot et al., 1969). The release of oxygen occurs in 1 ms, i.e. a 10 times shorter timeinterval than the overall bottleneck of the full electron transport chain (Emerson and Arnold, 1932). In 1970 Bessel Kok concluded that PSII contains an accumulator of four positive 'charges' that links the one-electron-progression of the photo-physical reaction centre to the four electron/proton-abstraction from two molecules of water to yield dioxygen (Kok et al., 1970). From there on, the catalytic centre has been conceived as stepwise progressing in a cycle over five oxidation states, S₀ to S₄. With S₁ being the most stable state in dark adapted material, oxygen is only produced after three steps of the cycle during the transition $S_4 \rightarrow S_0$. Two most important features of this four-stepped mechanism were soon recognized (see Renger (1977)): (a) the levelling of the energy demand of the four successive one-electron abstractions from water to match the fixed energy input provided by red quanta of light, and (b) the control (by sequestration and/or short live-time) of potentially harmful intermediates on the way from water to dioxygen ('cryptoradicals'; Renger, 1987). The cleavage of water to produce dioxygen was expected to liberate protons into the lumen of thylakoids. The stoichiometric H⁺/ e⁻-pattern over the four-stepped progression, $S_0 \rightarrow S_1 \rightarrow S_2 \rightarrow$ $S_3 \rightarrow S_4 + O_2$ was 1:0:1:2 (Förster *et al.*, 1981; Förster and Junge, 1985). For some time it was controversial between various labs (reviewed in Lavergne and Junge, 1993). Different patterns were observed in partially fragmented PSII-preparations. The seeming ambiguity has later been settled in favour of a proton release pattern of 1:0:1:2. It has required the discrimination by kinetic markers between transient electrostatic proton release/uptake at the membrane periphery (membrane Bohr effect) and chemical proton production from the catalytic core of PSII (Haumann and Junge, 1994) (see review in Junge et al. (2002)). It has been early noted that the release of two protons during the terminal, oxygen evolving reaction is kinetically biphasic with one proton appearing at about 100 µs and the other one at 1 ms half rise (Förster et al., 1981; Förster and Junge, 1985). The release of a first proton appears as a primer for the terminal reaction cascade involving four electron transfers (see Fig. 3 farther down).

148

149

150

151

152

153

154

155

156

157

158

159

160

161

162

163

164

165

166

167

168

169

170

171

172

173

174

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

George Cheniae established the necessity of manganese for oxy-128 gen evolution (Cheniae and Martin, 1967, 1970). Ken Sauer's lab in 129 Berkeley has focused on characterizing its function. Starting with 130 electron paramagnetic resonance (EPR) (Blankenship and Sauer, 131 1974), they turned with Mel Klein into X-ray spectroscopy 132 (Kirby et al., 1981a, 1981b) and, till today, into advanced X-ray dif-133 fraction (see below). When exciting dark adapted chloroplasts with 134 repetitive flashes of light they detected by EPR that some manga-135 nese was released into solution with period of four. It suggested 136 that manganese associated with PSII undergoes valence-changes 137 with a period of four (Wydrzynski and Sauer, 1980). From then 138 on it was accepted that manganese is involved in Kok's 'charge' 139 accumulator (reviewed in Sauer (1980)). When generating Kok's 140 state S₂ by excitation of dark adapted material with a single short 141 flash of light Charles Dismukes and Yona Siderer observed a mul-142 tiline EPR-signal which they attributed to a mixed-valence pair of 143 manganese, Mn(III)Mn(IV), in a dimeric or tetrameric Mn-cluster 144 (Dismukes and Siderer, 1981). For quite a while this attribution has 145 been the stronghold for the valence of the manganese cluster dur-146 ing the four transitions from S_0 to S_4 . 147

Enrichment and purification of PSII: P680, the Mn-cluster and tyrosine (Y_z)

Further progress required enrichment if not purification of PSII. Per Ake Albertson's lab pioneered the isolation of PSIIenriched membrane vesicles from spinach (Akerlund *et al.*, 1976; Andersson *et al.*, 1977). A number of consecutive studies demonstrated that the enrichment was facilitated by the intrinsic segregation between PSII in the appressed portions and PSI in the extended portions of stacked thylakoid membranes (Andersson and Anderson, 1980, 1988).

Purification of a manganese containing, oxygen evolving PSII was desirable. A first claim (Spector and Winget, 1979) was not generally reproducible and probably fraudulent. A great step forward were membrane fragments from spinach chloroplasts that revealed high rates of oxygen evolution and were virtually uncontaminated by PSI (Berthold et al., 1981). They were coined BBY-particles after their parents, Berthold, Babcock and Yocum. The absolute content of PSII was quantified via the previously found, and chemically still unidentified EPR-signal II (Babcock and Sauer, 1973), then coined Z, supposedly an electron donor to P680⁺ (Babcock and Sauer, 1975). In 1984 a fully competent PSII-core particle from spinach was isolated by Demetrios Ghanotakis with Gerald Babcock and Charles Yocum (Ghanotakis et al., 1984b). It contained four Mn-ions and one Z per 250 chlorophyll molecules. In addition to the hetero-dimeric core proteins of PSII, D1 and D2, and further proteins with antennae function (CP43, CP47) the complex contained peripheral proteins of 17, 23 and 33 kDa molecular mass. The removal of the 17 and 23 kDa proteins facilitated the access of external reductants to Z. It also impaired oxygen production which was however reconstituted by added Ca (Ghanotakis et al., 1984a, 1985a, 1985b).

At this time (i.e. before crystallization, BC) structural information on the mutual arrangement of the four Mn-ions, and a role of Ca in relation to the Mn-cluster was still lacking (for reviews see Yocum, 1991; Debus, 1992).

Kinetics of electron transfer between $P680^+$, Y_z and the Mn-cluster

P680 is photo-oxidized in less than 300 ps (Schatz *et al.*, 1987, 1988). In PSII-preparations with small size of the antennae system

the time of photon capture is shorter than in those with large antennae complement. In other words, PSII hosts an energetically 'shallow trap' where many transfers of excitation between the trap and its antennae complement precede the eventual charge separation (Schatz et al., 1988; Barter et al., 2001). P680⁺ is then reduced in nanoseconds. For lack of time resolution in the earlier work the reduction of P680⁺ was first detected in functionally impaired PSII where it was much delayed compared to normal (Döring et al., 1967). In likewise impaired material Gerald Babcock's group found that the slow reduction of P680⁺ kinetically matched the oxidation of the EPR-visible radical intermediate, Z (Boska et al., 1983). In fully competent, oxygen evolving PSII, on the other hand, P680⁺ is reduced in nanoseconds as later found in Horst Witt's lab. The rate of P680⁺-reduction depends on the state of Kok's 'charge accumulator' before photon-absorption. The half-rise time ranges around 20-40 ns when starting from S_0 and S_1 and rises to about 300 ns when starting from S_2 and S₃ (Brettel et al., 1984; Schlodder et al., 1984; Meyer et al., 1989). According to its UV-difference spectrum the electron donor to P680⁺ was identified as a tyrosine, and the rate of its oxidation in nanoseconds matched the one for the reduction of P680⁺ (Gerken et al., 1988). By site-directed mutagenesis the electron donor was eventually identified as tyrosine-161 on the D1-polypeptide of PSII (Debus et al., 1988a, 1988b). It has been coined Y_z since then. While Y_z is oxidized by P680⁺ in nanoseconds, it is reduced by the Mn-cluster in micro- to milliseconds. The half-rise time of Y_{z}^{+} -reduction and Mn-oxidation depends on the state S_i before photon absorption, namely 30, 110, 350 and 1300 µs when starting from S₀, S₁, S₂ and S₃, respectively (Dekker et al., 1984).

Around 1990 the three players of oxygen evolution, namely P680, Y_z and a $Mn_4(Ca)O_x$ -entity, had been identified, and the kinetic constants of electron transfer between them quantified. The challenge was to elucidate the mutual arrangement of P680/ Y_z/Mn_4Ca , the role of Ca, and, of course, how the dioxygen bond is formed.

Tentative structural models of the catalyst

The X-ray crystal structure of the reaction centre of purple bacteria (BRC), the first of a membrane protein ever, was published in 1985 (Deisenhofer *et al.*, 1985) (Nobel Prize to J. Deisenhofer, R. Huber and H. Michel in 1988). It was soon evident that its subunits L and M share sequence similarity with D1 and D2 of PSII (see Nitschke and Rutherford, 1991 for review). For quite a while the bacterial reaction centre served as scaffold for discussing the structure of PSII of oxygenic photosynthesis. Without any structural model of PSII proper, several labs tried to distil structural information from spectroscopic data.

Gary Brudvig's group compared the EPR-signals, attributable to state S₂ of the oxygen evolving centre, with EPR-signals of certain Fe₄S₄ proteins of known structure. Because of the similarity of their ferro- and antiferromagnetic exchange couplings they tentatively conceived a tetrameric Mn_4O_4 -cluster in a cubane-like arrangement as one possibility for PSII (Depaula *et al.*, 1986). Based thereupon they hypothesized a comprehensive reaction scheme (Brudvig and Crabtree, 1986). A 'naked' Mn_4O_4 -cluster (the protein was not considered!) was proposed to cycle over the five oxidation states S₀ to S₄, evolve oxygen during the transition S₄ \rightarrow S₀, release protons with the known stoichiometric pattern of 1:0:1:2, accommodate two water molecules to yield an Mn_4O_6 -cluster in an adamantane-like conformation, and 190

191

192

193

194

233

234

235

236

237

238

239

240

eventually to form the dioxygen bond (see Fig. 1 in Brudvig and Crabtree (1986)). Among three possible sets for the oxidation states of the four Mn ions the authors considered a very highvalence (HV) state, namely $Mn(IV)_3Mn(V)$ in S₄, as probable (Brudvig and Crabtree, 1986).

Mel Klein started investigations of the state of manganese in 195 photosynthesis by X-ray absorption spectroscopy (XAS) (Kirby 196 et al., 1981a, 1981b), Ken Sauer's group joined this field 197 (Goodin et al., 1984), and they went on together in Berkeley. 198 Using synchrotron radiation that was tuned to the K-edge of 199 manganese and calcium ions they studied the X-ray absorption 200 near edge structure (XANES), and the extended X-ray absorption 201 fine structure (EXAFS). The former gives information on valence 202 states, the latter yields the distance between the primarily excited 203 ion and its neighbours. A set of Mn-Mn distances was determined 204 (with very high (0.1 Å) precision) for the S₁- and S₂-states 205 (Yachandra et al., 1987a, 1987b). The most prominent Mn-Mn dis-206 tances were two times 2.7 and one time 3.3 Å. Their K-edge showed 207 shifts that were indicative of Mn-oxidation during $S_0 \rightarrow S_1$ and 208 $S_1 \rightarrow S_2$, and much less so during the transition $S_2 \rightarrow S_3$ (Roelofs 209 et al., 1996). Ligand centred as opposed to Mn-centred oxidation 210 was considered as a possibility (Messinger et al., 2001). The abso-211 lute valence of Mn₄ could then not be assigned unequivocally, 212 except for S₂ with probable configuration of (III, IV, IV, IV) 213 (Roelofs et al., 1996; Iuzzolino et al., 1998; Schiller et al., 1998). 214

The results of the Berkeley group were presented in a series of 215 reviews (Yachandra et al., 1993, 1996). The Mn-Mn distance of 216 2.7 Å was attributed to di-u-oxo-linked Mn-Mn and of 3.3 Å to 217 mono-µ-oxo-linked Mn-Mn, respectively, by comparison with 218 data on Mn-model complexes of known structure (e.g. Christou, 219 1989; Wieghardt, 1989). The Berkeley group proposed a 220 Mn₄-model of two di-µ-oxo-linked Mn-dimers that are linked by 221 one mono-µ-oxo bridge (the dimer of dimers model) (Yachandra 222 et al., 1993, 1996). However, a number of other structures for the 223 four oxygen-linked Mn-ions were also compatible with the same 224 set of Mn-Mn distances. In an attempt to resolve the structural 225 ambiguity Holger Dau studied linear dichroism of X-ray absorption 226 in oriented membrane fragments (Schiller et al., 1998) (for further 227 dichroism studies see Yano et al. (2006)). The additional informa-228 tion did not lead to an unequivocal structural model. The high pre-229 cision of the Mn–Mn distances as determined by EXAFS (<0.1 Å) 230 has, however, served to gauge the correctness of structural models 231 by X-ray diffraction and computational chemistry. 232

The proximity of Ca to the Mn-ions was eventually established (Latimer *et al.*, 1995), and the distance between strontium (the only functional substitute for calcium) and Mn was determined by EXAFS (Cinco *et al.*, 1998). Sr neighbours two Mn-ions at the same distance, 3.5 Å. From there on the catalytic centre of oxygen production has been conceived as a hetero-nuclear Mn₄Ca-cluster. It has been coined 'oxygen evolving complex', in short the OEC.

Various approaches to elucidate the structure and valence of 241 the Mn₄-cluster based on the multiline CW-EPR signal of state 242 S2 had produced largely different models (e.g. Ahrling and 243 Pace, 1995; Zheng and Dismukes, 1996) because of a highly 244 under-determined data set. Jeffrey Peloquin and David Britt 245 increased the number of observable parameters by pulsed EPR/ 246 ENDOR (Peloquin and Britt, 2001). A trinuclear Mn₃ with one 247 appended Mn (coined 'dangler') was better compatible with 248 their data than the Mn₄-cubanoid that was previously suggested. 249 They noted that 'each manganese has a unique coordination envi-250 ronment' and may serve 'a specific mechanistic purpose'. Based 251

on their EXAFS studies on the S_0 -state the Berkeley group considered a cluster with three di- μ -oxo-bridged Mn plus one outlier Mn as a possibility (Robblee *et al.*, 2002).

Tentative mechanistic models for catalysis

Several authors have speculated about the chemical mechanism of water oxidation. Most of them focused on the metal ions with one exception. Gerald Babcock emphasized an active role of Tyr¹⁶¹ of D1 (' Y_z '), namely as hydrogen acceptor for bound 'water' (Hoganson and Babcock, 1997; Tommos and Babcock, 1998). Following Lev Krishtalik (Krishtalik, 1989, 1990) they argued that a neutral tyrosyl radical abstracts hydrogen from bound water (or OH) because concerted transfer of H required less reorganization energy than the sequential transfer of an electron and proton. Babcock presented a detailed reaction scheme that was based on Brudvig's concept of a Mn₄-cubanoid. The question was whether or not tyrosine, when oxidized by P680⁺, was a neutral radical. Michael Haumann in the author's group investigated electrostatic relaxation versus chemical production of protons (Haumann and Junge, 1994) plus local electrochromism (Haumann et al., 1997b) in the reaction cascade between P680, Y_z and the Mn₄Ca-cluster. In intact PSII the oxidation of Y_z revealed strong local electrochromism and no proton release into the bulk, as if the oxidation of Y_z left a positive charge on tyrosine or closely nearby. It has been proposed that the tyrosineproton is shifted to a neighbouring base, B, to which Y_z is hydrogen-bonded (Ahlbrink et al., 1998; Junge et al., 2002). B was then identified as D1-His¹⁹⁰ (Hays et al., 1998). The sequestration of the tyrosine-proton in its vicinity holds only in intact PSII, it does not hold if smaller subunits of PSII or Ca are removed (see (Ahlbrink et al., 1998; Junge et al., 2002). The priming reaction for water oxidation is then conceived as follows: $P680^+Y_zHis \rightarrow P680Y_z\&HisH^+$.

Taken together it has implied that oxidized Y_z can act as an electron but not as a hydrogen acceptor to the catalytic metal cluster.

Vincent Pecoraro and his coworkers had studied the reactivity of synthetic manganese complexes. Based thereupon they proposed a mechanistic model for photosynthetic water oxidation (Pecoraro *et al.*, 1998). 'An essential feature of the model is the nucleophilic attack by calcium-ligated hydroxide on an electrophilic oxo group ligated to high-valent manganese to achieve the critical 0–0 bond formation step.'

The involvement of Mn(V) = oxo had been previously discussed by Johannes Messinger based on studies on slow water exchange (Messinger *et al.*, 1995). The proposed pivotal role of Mn(V) = oxo for the catalytic mechanism has been taken up by others, and it is highly debated till today (see below).

By density functional theory (DFT) Per Siegbahn and Robert Crabtree scrutinized possible mechanisms for the formation of an O–O bond (Siegbahn and Crabtree, 1999) in line with Babcock's and Pecoraro's proposals. By calculations of energy surfaces they found that hydrogen abstraction from Mn–OH by a neutral tyrosine radical probably could not *directly* produce a reactive, manganese bound oxyl. As a way out they proposed an indirect pathway involving Ca that is chelated to the manganese.

Crystal structures of PSII with the Mn₄CaO₅-cluster

Starting from his PhD-work in Göttingen (right after World War II) the mechanism of water oxidation to yield dioxygen has

252

253

254

255

256

257

258

259

260

261

262

263

264

265

266

267

268

269

270

271

272

273

274

275

276

277

278

279

280

281

282

283

284

285

286

287

288

289

290

291

292

293

294

295

296

297

298

299

remained Horst Witt's career-long preoccupation, his 'spröde Geliebte' (prudish beloved, see Junge and Rutherford (2007)). But it was only after retirement that he focused on crystallization of PSII, using a PSII complex from the thermophilic cyanobacterium Synechococcus sp. (Rögner et al., 1987). Witt joined forces with the crystallographer Wolfram Saenger, and, on their way to the structure of PSII, the Berlin group arrived first at crystals and a structural model of PSI 'of water splitting photosynthesis' (Witt et al., 1988). (Three years later it was improved to a resolution of 2.5 Å (Jordan et al., 2001).) Thereafter Athina Zouni characterized PSII-crystals capable of water oxidation (Zouni et al., 2000). The long desired first structural model of PSII was presented (Zouni et al., 2001). At a resolution of 3.8 Å it revealed a dimeric structure of PSII, the relative positions of its large subunits, the transmembrane helices, the position and orientation of cofactors (chlorophylls, Y_z, quinones, cytochromes), and the position and shape of Mn₄Ca as a whole, however without detail on the mutual arrangement of the metal ions.

Jian-Ren Shen and Nobuto Kamiya followed very closely. They characterized oxygen evolving crystals of PSII (Shen and Kamiya, 2000) and determined their structure at 3.7 Å resolution (Kamiya and Shen, 2003). The results were much alike those of the Berlin group with new features on ligands to the metal-cluster, and certain carotenoids. The inner structure of the metal-cluster was again presented as pear-shaped and without detail.

James Barber in London has not less been determined than the former two groups to solve the crystal structure of PSII (see Andersson (2005)). In 1998, in close collaboration with Werner Kühlbrandt and by electron crystallography of 2D crystals they derived a structural model of a PSII-subcomplex that was capable of photochemical electron transport but not of water oxidation. The location of four subunits and a total of 23 transmembrane helices was resolved at 8 Å (Rhee *et al.*, 1998). The structural similarity between the reaction centre of purple bacteria, PSI and PSII has been evident (see their Fig. 5). James Barber then turned to X-ray crystallography in collaboration with So Iwata, and they obtained a fully refined structural model of PSII at 3.5 Å resolution (Ferreira *et al.*, 2004). Over 5000 side chains of the heterodimeric protein complex were assigned.

Most important has been their model for the metal-cluster (see Fig. 2). In conceiving this far sighted structural model the authors relied on their own diffraction data, including the anomalous diffraction aiming at Mn (X-ray wavelength 1.89 Å) and Ca (2.25 Å), the Mn–Mn and Mn–Ca distances from EXAFS-studies of the Berkeley group (Yachandra *et al.*, 1987*b*), knowledge on μ -oxolinked Mn-model-clusters as previously discussed (Brudvig and Crabtree, 1989), and the proposed Mn₃ + Mn(dangler)-structure (Peloquin and Britt, 2001).

The Barber-model of the metal cluster had three Mn plus one 300 Ca forming a cubanoid and the fourth Mn dangling out. The ions 301 were bridged by four oxygen atoms and ligated by four water mol-302 ecules (though not clearly resolved). A bicarbonate was postulated 303 to bridge the Ca-ion and the dangling Mn. Four protein residues 304 ligated the Mn_4CaO_4 -cluster, with $Y_z = D1$ -Tyr¹⁶¹ and its hydro-gen-bonded partner, D1-His¹⁹⁰, in close vicinity to the former. In 305 306 essence, this model has set the path until today. One year later the 307 Berlin group presented a model at slightly higher resolution of 308 3 Å, where all carboxylate residues were now bi-dentate bridging. 309 Even the one that was terminally bound to one manganese in 310 Barber's model was now bridging two metals (Loll et al., 2005). 311 Again the resolution was not sufficient to resolve bridging oxygen 312 atoms and water. 313



Fig. 2. Left: The first structural model of the catalytic centre (Ferreira *et al.*, 2004). Right: Schematic representation of the $Mn_4CaO_5(H_2O)_4$ -complex with the numbering as used after the first model at high-resolution (Umena *et al.*, 2011).



Fig. 3. Tentative scheme for the cyclic stepping of the catalytic metal cluster over Kok's states $S_0 \rightarrow S_1 \rightarrow S_2 \rightarrow S_3 \rightarrow S_4$ and the formation of the dioxygen bond (Krewald *et al.*, 2016).

Johannes Messinger reviewed the status of agreements and diverging views up to 2004. He discussed two possible mechanisms for O–O bond formation, namely involving the dangler Mn and one Ca-bound water on the one hand, and involving a bridging oxygen and a dangler-bound water (Messinger, 2004).

Crystals exposed to a CW-X-ray beam from a synchrotron undergo radiation damage. By X-ray spectroscopy Holger Dau had detected dose-dependent Mn-reduction in PSII (Dau *et al.*, 1997). The larger Mn–Mn bond length in Barber's crystal structure as compared with the one inferred from EXAFS (Dau *et al.*, 2004) raised serious doubt on the valence-state of the Mn₄Ca-cluster in Barber's model. The Berlin group joined forces with the Berkeley group and they clearly demonstrated that X-ray exposure (doses by order of magnitude lower than causing loss of diffractivity) reduced Mn_4 from the expected valence of state S_1 (III,III,IV,IV) to (II,II,II,II) (Yano *et al.*, 2005). It explained why the Mn–Mn distances of the structural model were larger than very precisely determined by EXAFS. At the same time the valence-set of the four Mn-ions was determined by ⁵⁵Mn-pulse-NMR in Wolfgang Lubitz' lab. It was (III,III,III,IV) in S_0 and (III,V,IV,IV) in S_2 (Kulik *et al.*, 2005).

Soon thereafter Per Siegbahn took Barber's arrangement of the metals and their ligands (with modifications by the Berlin group) as the basis for modelling Mn_4CaO_x -clusters *in silico* (Siegbahn, 2006). By DFT and energy minimization a particular configuration of Mn_4CaO_5 was selected to represent the HV state S_4

from where the O-O bond is formed. The Mn-Mn bond lengths in the energy minimized structure were shorter than in Barber's. In this respect Siegbahn's modelling was predictive for radiation damage and not only post mortem descriptive as common in computational biochemistry/biophysics.

It was not astounding that, for a while, Barber's structure of the metal cluster was met with reservations, because it has been all but trivial, that the metal and ligand positions in PSII are the same if the valence of Mn_4 is (III,III,IV,IV), as supposed for S_1 , or (II,II,II,II), as after radiation damage.

Subsequent work on the atomic structure has been aimed at (a) minimization of radiation damage, (b) improved spatial resolution, (c) selective characterization of the five functional states S_0 to S_4 and (d) time resolution of structural transitions between them. The latter necessitates X-ray diffraction at room temperature.

The first high-resolution structure of PSII at 1.9 Å was published by Jian-Ren Shen and Nobuo Kamiya (Umena et al., 2011). Barber's cubanoid for Mn₃Ca with the dangling Mn (Ferreira et al., 2004), and the ligand structure of the Berlin group (Loll et al., 2005) were corroborated in essence. For the first time, the bridging oxygen atoms and four water molecules resolved, and former Mn₄CaO₄ became were clearly Mn₄CaO₅(H₂O)₄, as schematically illustrated in Fig. 2. Barber's numbering of the metal ions and oxygen atoms has been changed accordingly. The position of the dangler-Mn relative to the cubane was corrected by 3 Å, and Asp 170 was now a bidentate ligand (bridging Mn4 and Ca) and not monodentate, as before. This structural model was obtained by continuous synchrotron radiation (CW-XRD), and at cryogenic temperature. The Mn-Mn distances were still slightly longer than in both, EXAFS experiments and DFT calculations, and hence still indicative of some radiation damage.

A damage-free structure was desirable. X-ray diffraction with ultra-short pulses (typ. duration, <50 fs) of a free electron laser (XFEL) seemed to offer a way out. The pioneering study was published in 2012 by the joined groups of Berlin (i.a. Athina Zouni), Berkeley (i.a. Jan Kern, Junko Yano, Vittal Yachandra) and Umeå (Johannes Messinger). Very many stochastically oriented microcrystals of PSII were each exposed to a single X-ray shot to 'diffract before destruction' (Kern et al., 2012). It followed a series of such XFEL studies (e.g. Kern et al., 2013, 2014; Hellmich et al., 2014; Kupitz et al., 2014) carried out at room temperature and aiming at improved resolution.

Shen and his colleagues also conducted XFEL experiments. In contrast to the former group they used large crystals with volume-per-volume exposure at cryogenic temperature. In 2015 they presented a 'damage-free' structural model at a resolution of 1.95 Å (Suga et al., 2015). The Mn-Mn distances were shorter than previously obtained by conventional CW-XRD (Ferreira et al., 2004; Umena et al., 2011), and they were better compatible with those from EXFAS and DFT (for the remaining slight difference, see Fig. 1 in Siegbahn (2013b)). The 1.95 Å-structure has become the standard for the following discussions.

A long hedged dream of inorganic chemists came true when Chunxi Zhang and his colleagues synthesized a biomimetic analogue of nature's Mn₄Ca-cluster (Zhang et al., 2015).

The obvious challenge was now to determine the damage-free structure of the catalytic centre at room temperature, in its different oxidation states $(S_0, ..., S_4)$, and, if possible, time resolved.

In 2016 the Berkeley-Berlin team published a XFEL-study on structural differences between states S1 and S3 of the metal cluster, as function of the temperature, and with and without ammonia 376

377

378

379

380

381

382

383

384

385

386

387

388

389

390

391

392

393

394

395

396

397

398

399

400

401

402

403

404

405

406

407

408

409

410

411

412

413

414

415

419

420

421

422

423

424

425

426

427

428

429

430

431

432

433

434

435

436

437

bound (Young et al., 2016). With ammonia the authors mapped those water binding sites in the metal-cluster that are not involved in the formation of the O-O bond. While temperature affected some positions of transmembrane α -helices and redox-cofactors, the Mn₄CaO₅-cluster, proper, was practically invariant to temperature. At a resolution of 2.25 Å the authors did not detect differences between the dark state (S_1) and the 'two flashes advanced' state (mainly S_3) (Young *et al.*, 2016).

Photosystem II with the oxygen evolving complex (AC)

Mechanistic models for O-O bond formation

Adopting a chemical mechanism that had been proposed by V. Pecoraro and his colleagues (Pecoraro et al., 1998) James Barber speculated that the dioxygen-bond is formed by nucleophilic attack of a water ligand to the Ca-ion on a highly electrophilic oxo supposedly bound to the dangler-Mn(V) (Ferreira et al., 2004; Iwata and Barber, 2004). Barber has favoured this concept until today (Barber, 2016, 2017). His view is in line with earlier computational studies by P. Siegbahn (Siegbahn and Crabtree, 1999; Siegbahn, 2000), and it has been shared recently by others (McEvoy et al., 2005; Sproviero et al., 2006, 2008; Askerka et al., 2017).

After the first structural model of the metal-cluster had been published Per Siegbahn investigated by DFT the energy profile of various mechanistic models for the formation of the O-O bond. Calculations of the energy demand for an O-O-bonded intermediate in the Mn₄CaO₅-structure led him to reject the nucleophilic attack mechanism. He has since then proposed a radical attack mechanism involving a new bridging oxygen (O5) within an open cubanoid (coined oxyl-oxo-mechanism) (Siegbahn, 2006) that was not included in Barber's structural model. O5 resulted from 'predictive computational chemistry', and it was only later observed in diffraction experiments at higher resolution. Siegbahn has extended this work later on (Siegbahn, 2006, 2008a, 2017; Siegbahn and Blomberg, 2014).

Yu Guo and his coworkers embarked on similar DFT calculations and likewise favoured an oxo-oxyl mechanism involving the same bridging oxygen (O5) in an open cubanoid (Guo et al., 2017). On first inspection Guo's results are in line with 416 Siegbahn's, however a closer look reveals that their calculated 417 energy profile for the last transition $(S_3 \rightarrow S_0)$ violates the 418 experimentally established fact that oxygen liberation and Y_z⁺-reduction are both kinetically enslaved by the same rate limiting step. In his calculated energy profile these two reactions are separated by an energy notch, 12 kcal/mol deep (see their Fig. 4; Guo et al., 2017). Siegbahn's calculated energy profile on the other hand is compatible with the kinetic data.

The proponents of the leading concepts for the formation of the O-O bond, nucleophilic attack versus oxo-oxyl, have presented their diverging views in series of publications starting from 2004 (Ferreira et al., 2004) and 2006 (Siegbahn, 2006) until today. While James Barber most recently argued in favour of nucleophilic attack by analogy with the known mechanism of carbon dioxide dehydrogenase (Barber, 2017), Per Siegbahn (Siegbahn, 2017) repeated his previous (Siegbahn, 2006) exclusion of nucleophilic attack on energetic grounds.

In addition to the above two concepts for the formation of the O-O bonds several modified or even alternative mechanisms have been proposed (see e.g. Messinger, 2000; Gao et al., 2009; Yamanaka et al., 2011; Saito et al., 2012; Cox and Messinger, 2013).

Experimental tests of proposed mechanisms

The light driven stepping over the intermediate states of the 'charge accumulator' (Mn_4CaO_5) has been kinetically well characterized (for a comprehensive review, see Dau *et al.* (2012)). The eventually fourfold oxidized Y_2Mn_4Ca -entity reacts with two molecules of water (-derivatives) which involves the transfer of four electrons. The terminal four-electron-cascade is kinetically enslaved by a bottle neck of approximately 1 ms duration. Characterizing the short-lived intermediates is difficult if not impossible. The formation of a peroxide intermediate in the penultimate state S_3 has been in focus.

The valence of the four manganese ions in states S_i (i = 0-4)

A first stronghold for the assignment of oxidation states to the four Mn-atoms has been the EPR-multiline EPR-signal of the S2-state. It has been attributed to the interaction between Mn (III) and Mn(IV) (Dismukes and Siderer, 1981). Two valence configurations of the four Mn-ions are compatible with this notion, namely Mn(III)₃Mn(IV) and Mn(III)Mn(IV)₃ (de Paula and Brudvig, 1985). These assignments have been extended into two schemes for the S_i-series which are usually referred to as lowvalence (LV) and HV models. An enormous amount of work by different techniques had been devoted to discriminate between these models, for a while without reaching consent (reviewed in Krewald et al. (2015)). The comparison of NEXAFS-data with DFT-calculations supported the HV-model, although a few complications seemed to prevent an unequivocal attribution (Brena et al., 2012). Related to the above controversy has been the one over ligand-centred versus Mn-centred oxidation in the transition $S_2 \rightarrow S_3$. The former was advocated in Yachandra *et al.* (1996) and Messinger et al. (2001) and the latter in Iuzzolino et al., 1998; Dau et al., 2003; Siegbahn, 2009; Brena et al., 2012. ⁵⁵Mn-ENDOR studies on states S₀ and S₂ (Kulik et al., 2005) and XAS-studies on S₂ and S₃ (Dau et al., 2003) have established the HV concept for the four manganese ions, namely, $S_0 \rightarrow S_1 \rightarrow S_2 \rightarrow S_3 = (III, III,$ $\text{III,IV}) \rightarrow (\text{III,III,IV,IV}) \rightarrow (\text{III,IV,IV,IV}) \rightarrow (\text{IV,IV,IV,IV}).$

The oxidation of manganese during $S_2 \rightarrow S_3$ and the existence of a $Mn_4(IV)$ -state has been corroborated by X-ray emission spectroscopy (Zaharieva *et al.*, 2016*a*; Schuth *et al.*, 2018).

Two lines of computational studies have agreed on that the S_2 -state is interconvertible between two sub-states. In one state, coined S_2^B , the only trivalent ion is Mn4(III), the dangler outside of the cubane. In the other state, coined S_2^A , it is Mn1(III) within the cubane. Both these studies agreed in that only S_2^B can proceed to S_3 owing to the presence of a titratable ligand (Bovi *et al.*, 2013; Retegan *et al.*, 2016). While S_2^A is more stable than S_2^B the oxidation of Y_z inverts this situation such that S_2^B passes by proton-coupled electron transfer into state S_3 (Mn(IV)₄) (Narzi *et al.*, 2014).

Krewald *et al.* cast the available data, including their own on hyperfine coupling constants, into one consistent theoretical framework (Krewald *et al.*, 2015). They found that the most stable states, the open cubane structure of S₂ and the closed cubane of S₃, correspond to structures proposed earlier (based on DFT) by Siegbahn (Siegbahn, 2013*a*). In contrast to speculation in the literature their simulations yielded no evidence for ligand centred oxidation between S₂ and S₃. And they showed that only the HVmodel complies with the data but not the LV-model (Krewald *et al.*, 2015). In light of these results, state S₃ is to be conceived as Y_zMn₄(IV) or Y⁺_z^{Mn}Mn₃(IV)Mn(III) (Cox *et al.*, 2014; Retegan *et al.*, 2014), and state S₄ asY⁺_z^{Mn₄}(IV). The latter initiates the formation of the O–O bond. 438

450

451

452

453

454

455

456

457

458

459

460

461

462

463

464

465

466

467

468

469

470

471

472

473

474

475

476

477

478

479

480

481

482

483

484

485

486

487

488

489

490

491

492

493

494

495

496

497

498

499

The entry of water into the reaction cycle

The points of entry of solvent water into the reaction sequence of 439 oxygen evolution were at first characterized by mass spectroscopy 440 (Messinger et al., 1995; Hillier et al., 1998). The location of the 441 binding sites was then unknown. The µ-oxo-bridges were initially 442 considered as too strong for solvent exchange and catalytic rele-443 vance. Hydrogen exchange studies and ¹⁷O-NMR spectroscopy 444 has recently demonstrated that at least one µ-oxo bridge in the 445 metal cluster, either O4 or O5, is solvent exchangeable 446 (Rapatskiy et al., 2012). The latter μ -oxo might participate in 447 O-O bond formation as previously postulated by Siegbahn 448 (Siegbahn, 2006). 449

Dimitrios Pantazis and colleagues (Retegan et al., 2016) in Wolfgang Lubitz' lab in Mülheim discussed water exchange in the light of experiments on the Mn-coordination number in the S₃ state (Haumann et al., 2005) and of recent work on water exchange, when progressing through the S_i states (Suzuki *et al.*, 2008). One water molecule binds after oxygen release. It has been implied to form the di-µ-oxo bridge in the S₀-state (Krewald et al., 2015) and a fluxional bridge in the S₂-state (Bovi et al., 2013). The second water binds in the $S_2 \rightarrow S_3$ transition. It is probably not the reacting water in the next turnover (Suzuki et al., 2008; Cox and Messinger, 2013; Nilsson et al., 2014a). Pantazis proposed that W2 on Mn4, it pivots close to O5, might act as reaction partner in the formation of the O-O bond (Retegan et al., 2016). This view is however at odds with ammonia mapping of reactive water by the Berlin-Berkeley team which has let them to exclude W2 in this role (Young et al., 2016).

Christopher Kim and Richard Debus investigated the vibrational modes of bound water by FTIR-spectroscopy (Kim and Debus, 2017). The D-O-D bending-mode of one water molecule is altered upon functional substitution of Sr for Ca. This particular hydrogen-bonded water molecule is eliminated during the transition $S_2 \rightarrow S_3$. The authors proposed that this water molecule, probably W3, is deprotonated and moves to a position close to O5 as partner for O–O bond formation in the next transition ($S_3 \rightarrow$ S_4 - S_0). During $S_2 \rightarrow S_3$ the vacant water position on Ca is refilled by the incoming substrate water (Kim and Debus, 2017). Their view is compatible with the reorganization of hydrogen bonds as inferred from a large H/D-kinetic isotope effect on the reduction of Y₂& as monitored by UV- (Haumann et al., 1997a), X-ray-(Zaharieva et al., 2016b) and FTIR-spectroscopy (Sakamoto et al., 2017). Nicholas Cox and colleagues (Krewald et al., 2016) cast the available spectroscopic evidence into a spin-geared reaction scheme where the O-O bond is formed within the cubane. Their scheme is illustrated in Fig. 3.

The above spectroscopic results from Wolfgang Lubitz lab and other laboratories are in line with P. Siegbahn's oxyl-oxomechanism (Siegbahn, 2006, 2017) and at odds with J. Barber's nucleophilic attack mechanism (Ferreira *et al.*, 2004; Barber, 2017). They qualify alternative computational models (Kusunoki, 2007, 2011), in particular those based on LV of the Mn-cluster (Sproviero *et al.*, 2008; Gatt *et al.*, 2011; Petrie *et al.*, 2012; Li *et al.*, 2013). The challenge has been back to structural biology.

Structural evidence for/against proposed mechanisms of 0–0 bond formation

In 2016 the Berkeley–Berlin team published a XFEL-study on structural differences between states S_1 and S_3 of the metal cluster at room temperature (Young *et al.*, 2016). At a resolution of 2.25 Å no difference in electron density was detected between

the dark state (S_1) and the 'two flashes advanced' state (mainly S_3), as mentioned.

In 2017 Shen and coworkers in Japan reported structural differences of the metal-cluster between states S_1 and S_3 up to a resolution of 2.1 Å (Suga *et al.*, 2017). Their study was carried out at room temperature and under excitation with 10 fs pulses of a free electron laser. They found an 'apparent positive peak' in the difference Fourier map close to oxygen atom O5 which they interpreted as new oxygen, coined O6. O5 and O6 were taken as candidates for the formation of the O–O bond (Suga *et al.*, 2017) in line with Siegbahn's computational results (Siegbahn, 2006, 2008*b*, 2009). The authors discussed that Glu189, the only mono-dentate ligand to the cluster, had to move away from the cubane to accommodate O6.

When taken at face value this result could put an end to the major controversy over the mechanism of the O–O bond formation (nucleophilic attack *versus* oxyl-oxo).

At present, however, several features call for scrutiny: (a) the 'apparent positive peak' in Shen's difference map (interpreted as O6) is hardly elevated over noise and, at the given resolution, superimposed by electron density of Mn. (b) It is not yet clear to what extent it depends on particular software for the evaluation of raw data. (c) The purity of the dark state (100% S₁?) and of the '2-flash-advanced state' (45% S₃ in Shen's implies 55% S₂!) has been difficult to assess in crystals of PSII. (d) Spectroscopic evidence disfavours the presence of a peroxide intermediate in state S₃. (e) The supposedly 'moving' residue Glu189 has been point mutated into Arg, Lys and Gln without influence on the rates of various electron transfer reactions at the donor side of PSII (Clausen *et al.*, 2001).

The Berlin-Berkeley group has advanced the search technique for intermediates of the O-O bond by combining serial femtosecond X-ray crystallography with simultaneous X-ray spectroscopy under multi-flash visible laser excitation (Kern et al., 2013). In principle, this technique allows time resolution of structural transitions by variation of the time delays between the X-ray and the visible flashes. The major challenge is to obtain the necessary spatial resolution (<2 Å) to reveal the expected displacement of water (oxygen) molecules in 100 μ s during the transition from S₂ towards S₃ and onwards via 'S₄' to S₀ in a millisecond. Recently, this team has reported the structure of all states of Kok's cycle at 2.04-2.08 Å resolution including two transient states during $S_2 \rightarrow S_3$ at 150 and 400 µs after excitation by a flash of visible light (Kern et al., 2018). Their data exclude the formation of a peroxide intermediate already in state S₃. In S₃ they found the binding of a new 'Ox' (a water derivative) in a position between Mn1 and Ca. In the next step, $S_3 \rightarrow S_4 - S_0$ it could either form an O-O bond with O5 or replace O5 after formation of such a bond between O5 and another Ox. These results are in perfect agreement with the above cited results obtained by X-ray and magnetic resonance spectroscopy. There is no evidence that a peroxide intermediate is already formed in state S₃. But it is expected that one intra-cubanoid oxo-group, namely O5, is involved in the formation of the O–O bond in the next step, $S_3 \rightarrow S_4$ – S_0 .

Perspective for a rigorous resolution of the molecular mechanism

The rigorous resolution (in space and time) of the four oneelectron steps and the eventual four-electron reaction cascade of this pivotal reaction for life on earth is complicated by two features: (a) the great stability of the (Mn_4CaO_5cage)-structure and (b) the seeming kinetic enslaving of six partial reactions by one particular step.

(a) The Mn_4CaO_5 -cluster is very stable, and its expected structural changes during the terminal reaction cascade are probably subtle. Even gross valence changes of Mn like those caused by radiation damage in CW-X-ray diffraction (Dau *et al.*, 1997; Yano *et al.*, 2005) increase the bond length between metal ions only by small amount (see Siegbahn (2009)). This is why mechanistic insight can hardly be expected from tracking the motion of the metal ions proper.

As already mentioned, the effects of radiation damage have been minimized by either extremely low dose in conventional synchrotron X-ray diffraction (Tanaka *et al.*, 2017) and by femtosecond irradiation in XFEL-diffraction experiments (Kern *et al.*, 2014; Tran *et al.*, 2014; Suga *et al.*, 2015). Another type of radiation damage, namely a 'coulomb-explosion' of the metal cluster in some 10 fs has only recently been considered. MD-simulations have revealed that it is minimized if the X-ray pulse length is shorter than 10 fs (Amin *et al.*, 2017, 2016).

It came as a surprise that not only the metal-cluster is extremely stable but its protein cage as well. The Berlin group deleted the metal-cluster from PSII, and they found the protein cage very little altered, as if waiting for the insertion of di- μ -oxobridged pairs of manganese (Zhang *et al.*, 2017).

(b) The key to fully understand the mechanism of water oxidation are the trajectories of water, hydroxyl, oxygen, proton and electron. The motion of the former three might be tracked in the near future by time-resolved XFEL-crystallography at <2 Å resolution. Spatial tracking of protons (and electrons) is however not yet in sight. Here one is restricted to kinetic evidence based on spectroscopic data. Here, the kinetic enslaving of six partial reactions by one particular step represents a major challenge as illustrated in Fig. 4.

If the valence of Mn₄ in state S₂ is taken as a reference, namely (IV, IV, IV, III), the penultimate state before oxygen release possibly is: $S_3 = [Mn(IV)_4CaO_5(H_2O)_4Ligands]^+$. The next photoexcitation creates the sequence of events that is illustrated in Fig. 4. Oxygen is released plus two protons and one water molecule is taken up. The release of two protons is kinetically bipartite, a first step in the time range of 100 µs is followed by a slower step at 1 ms (Förster et al., 1981). The fast release of one proton seems to be the priming event for the subsequent cascade of six reactions which are kinetically enslaved by one and the same bottle neck of 1 ms half-duration, namely (1) the transfer of one electron to Y_{z}^{*} HisH⁺, (2) the reduction of three manganese ions, (3) the release of a second proton, (4) the release of di-oxygen and (5) the uptake of one molecule of water. The exact sequence of these events is subject to ongoing research. Michael Haumann and Holger Dau ventured into time-resolved calorimetry using photo-thermal beam deflection (Krivanek et al., 2008; Klauss et al., 2009). They obtained a wealth of signal transients that were interpreted to show the release of one proton first, oxygen release and water intake second and the release of another proton third (see Fig. 8 in Klauss et al. (2015)). It seems worthwhile to take up such studies with certain mutant-PSII were oxygen release is dramatically slowed down (Hundelt et al., 1998; Clausen et al., 2004; Nilsson et al., 2014b).

Energetics of oxygen production by PSII

Aiming at the stabilization of transient intermediates of the terminal reaction cascade PSII was exposed to high oxygen pressure 500

501

502

503

504

505

506

507

508

509

510

511

512

513

514

515

516

517

518

519

520

521

522

523

524

525

526

527

528

529

530

531

532

533

534

535

536

537

538

539

540

541

542

543

544

545

546

547

548

549

550

551

552

553

554

555

556

557

558

559

560



Fig. 4. The terminal reaction cascade of oxygen formation (see the text).

(Clausen and Junge, 2004). Certain UV-absorption transients that clearly showed a phase of ~1 ms half-time during $S_3 \rightarrow S_4 - S_0$ were monitored. These transients had been previously attributed to the reduction of the manganese cluster (Dekker et al., 1984). If the oxygen pressure was increased 100-fold over atmospheric this phase was virtually eliminated. Half-inhibition was reached at \approx 10-fold increase. It suggested a rather low driving force for photosynthetic oxygen production, implying 'little leeway for photosynthetic organisms to push the atmospheric oxygen concentration much above the present level' (Clausen and Junge, 2004) (for possible ecological and geochemical impact see Raven and Larkum (2007)). At first this observation seemed to be corroborated by another technique, delayed chlorophyllluminescence (Clausen et al., 2005b). Later however, no stalling effect of increased oxygen pressure was detected by (a) timeresolved X-ray absorption aiming at the K-edge of Mn (Haumann et al., 2008), (b) chlorophyll-fluorescence (Kolling et al., 2009) and (c) direct detection by membrane-inlet mass spectroscopy of ¹⁸O₂ produced from H₂¹⁸O against a background of pressurized ¹⁶O₂ (Shevela et al., 2011). In the latter experiments the oxygen pressure was raised up to 20 bar without any indication for stalling of dioxygen liberation. It implies that the driving force is at least 200 meV. Recently this number was driven up by another approach. In one cyanobacterial mutant-PSII the S₀-state is stabilized for days. Johannes Messinger and his colleagues incubated this mutant for up to 3 days in H₂¹⁶O and ¹⁸O₂ without detecting the formation of any amount of 16,18O2 (Nilsson et al., 2016). They concluded that the driving force for dioxygen release during $S_3 \rightarrow S_4 - S_0$ must be greater than 400 meV. They understood this large driving force in terms of Lev Krishtalik's pioneering discussion of the energetics of photosynthetic oxygen evolution in terms of thermodynamic principles (Krishtalik, 1989, 1990). Jérome Lavergne in Nilsson et al. (2016) attributed the experimentally established high driving force to the large entropic effect of dilution of bound oxygen into the solution (see free energy profile in Fig. 6 of Nilsson et al. (2016)). Their conclusion was that the protein has no grip on this 'extra enzyme' contribution to the driving force.

In oxygenic photosynthesis the entropy of dilution favours the release of freshly formed and still bound dioxygen. What about cell respiration where cytochrome oxidase has to overcome this huge entropic barrier?

The 'equilibrium constant' for oxygen in cytochrome oxidase was studied by Marten Wikström's group (Krab et al., 2011).



Fig. 5. The efficiency of solar energy conversion by oxygenic photosynthesis as a function of time (see the text).

The authors determined the apparent $K_m(O_2)$ for steady turnover of the enzyme, at neutral pH and in the absence of protonmotive force, $K_m(O_2) = 25$ nM (see their Fig. 4A). The free energy of oxygen binding from O₂-saturated water is in the order of -450 mV, very strong indeed, when considering the more than the +300 mV of Krishtalik's entropic contribution to oxygen dilution (see discussion in Nilsson *et al.* (2014*b*)).

Oxygenic photosynthesis and a sustainable future of mankind

Oxygenic photosynthesis has produced the oxygen we breathe and the biomass serving as food, feed, fibre and fuel, as mentioned. Its driving force is sunlight. The mean power of sunlight at the surface of earth is huge, 124×10^{15} W = 124 PW. Only a very small fraction thereof is captured by photosynthetic organisms, 89×10^{12} W = 89 TW, about half and half on land and in the sea (Field *et al.*, 1998; Falkowski *et al.*, 2000). The low proportion of solar energy capture is partially owed to unfavourable climate, substrate, seasons and the day–night cycle. Another loss of energy capture is intrinsic to the physico-chemical mechanism of photosynthesis, as described above. The solar energy conversion efficiency of photosynthesis is limited even under the most favourable conditions of agriculture, and even more so, if one aims at processed products like biofuels (see Fig. 5).

In the range of microseconds after absorption of a quantum of light the efficiency reaches up to 20% if related to the solar spectrum at the sea level (right scale in Fig. 5). It is about 80% under 607 excitation with red light (see right scale in Fig. 5). These figures 608 refer to a thin canopy (for a thick canopy, see Dau and 609 Zaharieva (2009)). Therewith the efficiency matches the one of 610 photovoltaic cells (Blankenship et al., 2011). The initial drop (to 611 20%) is mainly caused by (a) limited use of the full solar spectrum 612 (~50%) and (b) internal energy conversion ('blue \rightarrow red') within 613 chlorophyll. In the test tube and within the time range of 614 100 ms to yield NADPH the efficiency drops by another factor 615 of two. William Rutherford has coined this drop as 'sacrificing 616 energy efficiency for directionality' (Rutherford et al., 2012). In 617 other words, wasteful back-reactions are overcome by high 618 speed of the forward reaction which requires an energy drop. 619 On the physiological time scale (h) the efficiency drops further 620 from the best vegetative month of the year to the average over a 621 full year. The maximum yearly averaged efficiency at the given 622 level of atmospheric CO₂ (380 ppm in 2010) has been estimated 623

Fig. 4 - Colour online



Fig. 6. Left: Distribution over taxa of the biomass on earth in terms of Gt of fixed carbon (Bar-On *et al.*, 2018). Right: Solar energy fixation by global photosynthesis in TW (green) and energy consumption of mankind, both metabolically (blue) and technically (brown).

by Don Ort's group as 4.6% for C3- and 6% for C4-plants (Zhu et al., 2008). The efficiency of crops in the field is mostly lower. Take Brazil's large scale energy farming as an example. In 2010 the area specific yield of sugarcane was 80 t_{dwt} ha⁻¹ yr⁻¹. It implies a solar energy conversion efficiency of about 2%. The consecutive conversion of sugarcane into bio-ethanol yielded 63001 ha⁻¹ yr⁻¹ EthOH, equivalent of an overall energy efficiency of about 0.2%. Slops and slurry were utilized in the process, and the fossil energy input was not counted in this estimate. Various well-to-wheels analyses for biofuel production in the USA and in Europe (e.g. by the JRC in 2012: ISBN 978-9279-21395-3, ISSN 1831-9424, EUR 24952 EN, doi:10.2788/79018 and Torchio and Santarelli, 2010; Wang et al., 2012; Orsi et al., 2016) came to the same conclusion. The overall energy yield of biofuel production is at best 10% of the energy content of the biomass if not being negative (i.e. requiring higher fossil fuel input than biofuel output). Energy farming for biofuels is a questionable option, at least for densely populated countries that cannot waste arable and/or ecologically valuable land.

In a recent meta-study Ron Milo and his colleagues reviewed the distribution over taxa of the biomass on earth (Bar-On et al., 2018). Figure 6 (left) shows their result. In terms of fixed carbon, plants (83%) dominate over bacteria (13%) and animals (0.4%). Homo sapiens account for less than 0.01% of total. (By the way, the total amount of fixed carbon in sapiens is by one order of magnitude greater than that of all wild mammals taken together.) Confrontation of the biomass distribution over taxa (Fig. 6, left) with the energy provision by photosynthesis (Fig. 6, right, green) and the global primary energy consumption of mankind (Fig. 6, right, brown), shows that mankind, summed over all nations, uses 22% of what is provided by photosynthesis. Mankind's metabolic energy intake (Fig. 6, right, blue) is much less than the global energy consumption by technical civilization (Fig. 6, right, brown). It is noteworthy that the energy consumption per capita in industrialized countries like Japan and Germany is twice the global average, and in the USA it is more than four times greater. In other words, if the present global population cared to live in the style of USA citizens, they had to use the whole biomass production at almost 100% energy conversion efficiency, an absurd scenario.

Accordingly the engineering of plants and algae for higher efficiency (see e.g. Ort *et al.* (2015); South *et al.* (2018)) should be directed towards improved supply of food, feed, fibre and platform chemicals rather than of fuel. Having reached the Anthropocene, mankind cannot rely on oxygenic photosynthesis to satisfy its energy needs.

Acknowledgements. The writing of this paper was triggered during a meeting in Singapore in honour of Bertil Andersson, the past president of NTU. So it is for you, Bertil.

I am very grateful for constructive criticism by my friends and colleagues James Barber, Holger Dau, Michael Haumann, Johannes Messinger, Per Siegbahn, Mårten Wikström and Athina Zouni.

References

- Ahlbrink R, Haumann M, Cherepanov D, Bogershausen O, Mulkidjanian A and Junge W (1998) Function of tyrosine Z in water oxidation by photosystem II: electrostatical promotor instead of hydrogen abstractor. *Biochemistry* 37, 1131–1142.
- Ahrling KA and Pace RJ (1995) Simulation of the S2 state multiline electron paramagnetic resonance signal of photosystem II: a multifrequency approach. *Biophysical Journal* 68, 2081–2090.
- Akerlund HE, Andersson B and Albertsson PA (1976) Isolation of photosystem-ii enriched membrane-vesicles from spinach-chloroplasts by phase partition. *Biochimica et Biophysica Acta* **449**, 525–535.
- Amin M, Badawi A and Obayya SS (2016) Radiation damage in XFEL: case study from the oxygen-evolving complex of photosystem II. Scientific Reports 6:36492, 1–6.
- Amin M, Askerka M, Batista VS, Brudvig GW and Gunner MR (2017) X-ray free electron laser radiation damage through the s-state cycle of the oxygen-evolving complex of photosystem II. *Journal of Physical Chemistry B* 121, 9382–9388.

624

625

626 627

628

629 630 631

632

633 634 635

636

637 638

639

640 641 642

643

644

645

646

647 648 649

650

651

652

653

654

655

656

657

658

659

660

661

662

663

664

665

666

667

668

669

670

671

672

673

674

675

676

677

678

679

680

681

682

683

684

690

691

692

693

694

695

696

697

698

699

700

701

702

703

704

705

706

707

708

709

710

711

712

713

714

715

716

717

718

719

720

721

722

723

724

725

726

727

728

729

730

731

733

734

735

736

737

738

739

740

741

742

743

744

745

746

- Anderson JM and Andersson B (1988) The dynamic photosynthetic membrane and regulation of solar energy conversion. Trends in Biochemical Sciences 13, 351-355.
- Andersson B (2005) Introduction to the special issue dedicated to James Barber. Photochemical & Photobiological Sciences 4, 930-931.
- Andersson B and Anderson JM (1980) Lateral heterogeneity in the distribution of chlorophyll-protein complexes of the thylakoid membranes of spinach chloroplasts. Biochimica et Biophysica Acta 593, 427-440.
- Andersson B, Akerlund HE and Albertsson PA (1977) Light-induced reversible proton extrusion by spinach-chloroplast photosystem 2 vesicles isolated by phase partition. FEBS Letters 77, 141-145.
- Askerka M, Brudvig GW and Batista VS (2017) The O-2-evolving complex of photosystem II: recent insights from Quantum Mechanics/Molecular Mechanics (QM/MM), Extended X-ray Absorption Fine Structure (EXAFS), and femtosecond X-ray crystallography data. Accounts of Chemical Research 50, 41-48.
- Babcock GT and Sauer K (1973) Electron paramagnetic resonance signal II in spinach chloroplasts. I. Kinetic analysis for untreated chloroplasts. Biochimica et Biophysica Acta 325, 483-503.
- Babcock GT and Sauer K (1975) Rapid component of electron-paramagnetic resonance signal II - candidate for physiological donor to photosystem-II in spinach-chloroplasts. Biochimica et Biophysica Acta 376, 329-344.
- Bar-On YM, Phillips R and Milo R (2018) The biomass distribution on Earth. Proceedings of the National Academy of Sciences of the United States of America 115, 6506-6511.
- Barber J (2016) Photosystem II: the water splitting enzyme of photosynthesis and the origin of oxygen in our atmosphere. Quarterly Reviews of Biophysics 49. 1-20.
- Barber J (2017) A mechanism for water splitting and oxygen production in photosynthesis. Nature Plants 3:17041, 1-5.
- Barter LMC, Bianchietti M, Jeans C, Schilstra MJ, Hankamer B, Diner BA, Barber J, Durrant JR and Klug DR (2001) Relationship between excitation energy transfer, trapping, and antenna size in photosystem II. Biochemistry 40, 4026-4034.
- Bekker A, Holland HD, Wang PL, Rumble D, Stein HJ, Hannah JL, Coetzee LL and Beukes NJ (2004) Dating the rise of atmospheric oxygen. Nature 427, 117-120.
- Berthold DA, Babcock GT and Yocum CF (1981) A highly resolved, oxygenevolving photosystem-ii preparation from spinach thylakoid membranes electron-paramagnetic-res and electron-transport properties. FEBS Letters 134, 231-234.
- Blankenship RE (2014) Molecular Mechanisms of Photosynthesis. Chichester: Wiley Blackwell.
- Blankenship RE and Sauer K (1974) Manganese in photosynthetic oxygen evolution. 1. Electron-paramagnetic resonance study of environment of manganese in Tris-washed chloroplasts. Biochimica et Biophysica Acta 357, 252-266.
- Blankenship RE, Tiede DM, Barber J, Brudvig GW, Fleming G, Ghirardi M, Gunner MR, Junge W, Kramer DM, Melis A, Moore TA, Moser CC, Nocera DG, Nozik AJ, Ort DR, Parson WW, Prince RC and Sayre RT (2011) Comparing photosynthetic and photovoltaic efficiencies and recognizing the potential for improvement. Science 332, 805-809.
- Boardman NK and Anderson JM (1964) Isolation from spinach chloroplasts of particles containing different proportions of chlorophyll alpha + chlorophyll beta + their possible role in light reactions of photosynthesis. Nature 203, 166.
- Boska M, Sauer K, Buttner W and Babcock GT (1983) Similarity of EPR signal II f rise and P-680 + decay kinetics in Tris-washed chloroplast photosystem II preparations as a function of pH. Biochimica et Biophysica Acta 722, 327-330.
- Bovi D, Narzi D and Guidoni L (2013) The S-2 state of the oxygen-evolving complex of photosystem II explored by QM/MM dynamics: spin surfaces and metastable states suggest a reaction path towards the S-3 state. Angewandte Chemie-International Edition 52, 11744-11749.
- Brena B, Siegbahn PEM and Agren H (2012) Modeling near-edge fine structure X-ray spectra of the manganese catalytic site for water oxidation in photosystem II. Journal of the American Chemical Society 134, 17157-17167.

- Brettel K. Schlodder E and Witt HT (1984) Nanosecond reduction kinetics of 686 photooxidized chlorophyll-aII (P 680) in single flashes as a probe for the 687 electron pathway, H+-release and charge accumulation in the O 2 -evolving 688 complex. Biochimica et Biophysica Acta 766, 403-415. 689
- Brinkert K, De Causmaecker S, Krieger-Liszkay A, Fantuzzi A and Rutherford AW (2016) Bicarbonate-induced redox tuning in photosystem II for regulation and protection. Proceedings of the National Academy of Sciences of the United States of America 113, 12144-12149.
- Brudvig GW and Crabtree RH (1986) Mechanism for photosynthetic O-2 evolution. Proceedings of the National Academy of Sciences of the United States of America 83, 4586–4588.
- Brudvig GW and Crabtree RH (1989) Bioinorganic chemistry of manganese related to photosynthetic oxygen evolution. Progress in Inorganic Chemistry 37, 99-142.
- Burk D, Schade AL, Hunter J and Warburg O (1951) 3-Vessel and one-vessel manometric techniques for measuring Co2 And O-2 gas exchanges in respiration and photosynthesis. Symposia of the Society for Experimental Biology 5, 312-335.
- Cardona T, Murray JW and Rutherford AW (2015) Origin and evolution of water oxidation before the last common ancestor of the cyanobacteria. Molecular Biology and Evolution 32, 1310-1328.
- Cheniae GM and Martin IF (1967) Photoreactivation of manganese catalyst in photosynthetic oxygen evolution. Biochemical and Biophysical Research Communications 28, 89.
- Cheniae GM and Martin IF (1970) Sites of function of manganese within photosystem-II - roles in O2 evolution and system-II. Biochimica et Biophysica Acta 197, 219.
- Christou G (1989) Manganese carboxylate chemistry and its biological relevance. Accounts of Chemical Research 22, 328-335.
- Cinco RM, Robblee JH, Rompel A, Fernandez C, Yachandra VK, Sauer K and Klein MP (1998) Strontium EXAFS reveals the proximity of calcium to the manganese cluster of oxygen-evolving photosystem II. Journal of Physical Chemistry B 102, 8248-8256.
- Clausen J and Junge W (2004) Detection of an intermediate of photosynthetic water oxidation. Nature 430, 480-483.
- Clausen J, Winkler S, Hays AMA, Hundelt M, Debus RJ and Junge W (2001) Photosynthetic water oxidation: mutations of D1-Glu189K, R and Q of Synechocystis sp. PCC6803 are without any influence on electron transfer rates at the donor side of photosystem II. Biochimica et Biophysica Acta 1506, 224-235.
- Clausen J, Debus RJ and Junge W (2004) Time-resolved oxygen production by PSII: chasing chemical intermediates. Biochimica et Biophysica Acta 1655, 184-194.
- Clausen J, Beckmann K, Junge W and Messinger J (2005a) Evidence that bicarbonate is not the substrate in photosynthetic oxygen evolution. Plant Physiology 139, 1444–1450.
- Clausen J, Junge W, Dau H and Haumann M (2005b) Photosynthetic water oxidation at high O2 backpressure monitored by delayed chlorophyll fluorescence. Biochemistry 44, 12775-12779.
- Cox N and Messinger J (2013) Reflections on substrate water and dioxygen formation. Biochimica et Biophysica Acta-Bioenergetics 1827, 1020-1030. 732
- Cox N, Retegan M, Neese F, Pantazis DA, Boussac A and Lubitz W (2014) Electronic structure of the oxygenevolving complex in photosystem II prior to O-O bond formation. Science 345, 804-808.
- Dau H and Zaharieva I (2009) Principles, efficiency, and blueprint character of solar-energy conversion in photosynthetic water oxidation. Accounts of Chemical Research 42, 1861-1870.
- Dau H, Dittmer J, Iuzzolino L, Schiller H, Dorner W, Heinze I, Sole VA and Nolting HF (1997) X-ray absorption linear dichroism spectroscopy (XALDS) on the photosystem II manganese complex: radiation damage and S-1-state K-edge spectra. Journal De Physique IV 7, 607-610.
- Dau H, Liebisch P and Haumann M (2003) X-ray absorption spectroscopy to analyze nuclear geometry and electronic structure of biological metal centers - potential and questions examined with special focus on the tetranuclear manganese complex of oxygenic photosynthesis. Analytical and Bioanalytical Chemistry 376, 562-583.
- Dau H, Liebisch P and Haumann M (2004) The structure of the manganese complex of photosystem II in its dark-stable S-1-state-EXAFS results in

relation to recent crystallographic data. *Physical Chemistry Chemical Physics* 6, 4781–4792.

- Dau H, Zaharieva I and Haumann M (2012) Recent developments in research on water oxidation by photosystem II. *Current Opinion in Chemical Biology* 16, 3–10.
- **De Paula JC and Brudvig GW** (1985) Magnetic properties of manganese in the photosynthetic O2-evolving complex. *Journal of the American Chemical Society* **107**, 2643–2648.
- Debus RJ (1992) The manganese and calcium ions of photosynthetic oxygen evolution. *Biochimica et Biophysica Acta* 1102, 269–352.
- **Debus RJ, Barry BA, Babcock GT and Mcintosh L** (1988*a*) Site-directed mutagenesis identifies a tyrosine radical involved in the photosynthetic oxygen-evolving system. *Proceedings of the National Academy of Sciences of the United States of America* **85**, 427–430.
- **Debus RJ, Barry BA, Sithole I, Babcock GT and Mcintosh L** (1988*b*) Directed mutagenesis indicates that the donor to P-680+ in photosystem-II is tyrosine-161 of the D1 polypeptide. *Biochemistry* **27**, 9071–9074.
- Deisenhofer J, Epp O, Miki K, Huber R and Michel H (1985) Structure of the protein subunits in the photosynthetic reaction centre of *Rhodopseudomonas viridis* at 3 resolution. *Nature* **318**, 618.
- Dekker JP, Plijter JJ, Ouwehand L and Van Gorkom HJ (1984) Kinetics of manganese redox transitions in the oxygen evolving apparatus of photosynthesis. *Biochimica et Biophysica Acta* **767**, 176–179.
- Depaula JC, Beck WF and Brudvig GW (1986) Magnetic-properties of manganese in the photosynthetic O-2-evolving complex. 2. Evidence for a manganese tetramer. *Journal of the American Chemical Society* 108, 4002–4009.
- **Dismukes GC and Siderer Y** (1981) Intermediates of a polynuclear manganese center involved in photosynthetic oxidation of water. *Proceedings of the National Academy of Sciences of the United States of America* **78**, 274–278.
- **Döring G, Stiehl HH and Witt HT** (1967) A second chlorophyll reaction in the electron chain of photosynthesis – registration by repetitive excitation technique. *Zeitschrift fur Naturforschung Part B-Chemie Biochemie Biophysik Biologie und Verwandten Gebiete, B* **22**, 639–644.
- **Duysens LNM, Huiskamp WJ, Vos JJ and Vanderhart JM** (1956) Reversible changes in bacteriochlorophyll in purple bacteria upon illumination. *Biochimica et Biophysica Acta* **19**, 188–190.
- Duysens LN, Kamp BM and Amesz J (1961) Two photochemical systems in photosynthesis. *Nature* **190**, 510.
- Emerson R and Arnold W (1932) The photochemical reaction in photosynthesis. *The Journal of General Physiology* 16, 191–205.
- Engelmann TW (1881) xxxx. Botanische Zeitung, 39.
- Falkowski P, Scholes RJ, Boyle E, Canadell J, Canfield D, Elser J, Gruber N, Hibbard K, Hogberg P, Linder S, Mackenzie FT, Moore III B, Pedersen T, Rosenthal Y, Seitzinger S, Smetacek V and Steffen W (2000) The global carbon cycle: a test of our knowledge of earth as a system. *Science* **290**, 291–296.
- Ferreira KN, Iverson TM, Maghlaoui K, Barber J and Iwata S (2004) Architecture of the photosynthetic oxygen-evolving center. *Science* **303**, 1831–1838.
- Field CB, Behrenfeld MJ, Randerson JT and Falkowski P (1998) Primary production of the biosphere: integrating terrestrial and oceanic components. *Science* **281**, 237–240.
- Förster V and Junge W (1985) Stoichiometry and kinetics of proton release upon photosynthetic water oxidation. *Photochemistry and Photobiology* 41, 183–190.
- Förster V, Hong YQ and Junge W (1981) Electron-transfer and proton pumping under excitation of dark-adapted chloroplasts with flashes of light. *Biochimica et Biophysica Acta* 638, 141–152.
- Gao Y, Akermark T, Liu JH, Sun LC and Akermark B (2009) Nucleophilic attack of hydroxide on a Mn-V oxo complex: a model of the O–O bond formation in the oxygen evolving complex of photosystem II. *Journal of the American Chemical Society* **131**, 8726.
- Gatt P, Stranger R and Pace RJ (2011) Application of computational chemistry to understanding the structure and mechanism of the Mn catalytic site in photosystem II – a review. *Journal of Photochemistry and Photobiology B-Biology* **104**, 80–93.

- Gerken S, Brettel K, Schlodder E and Witt HT (1988) Optical characterization of the immediate electron donor to chlorophyll aII+ in oxygenevolving photosystem II complexes. Tyrosine as possible electron carrier between chlorophyll aII and the water-oxidizing manganese complex. *FEBS Letters* 237, 69–75.
 748

 FEBS Letters
 237, 69–75.
 752
- **Ghanotakis DF, Babcock GT and Yocum CF** (1984*a*) Calcium reconstitutes high-rates of oxygen evolution in polypeptide depleted photosystem-II preparations. *FEBS Letters* **167**, 127–130.
- **Ghanotakis DF, Babcock GT and Yocum CF** (1984*b*) Structural and catalytic properties of the oxygen-evolving complex. Correlation of polypeptide and manganese release with the behavior of Z+ in chloroplasts and a highly resolved preparation of the PS II complex. *Biochimica et Biophysica Acta* **765**, 388–398.
- **Ghanotakis DF, Babcock GT and Yocum CF** (1985*a*) On the role of watersoluble polypeptides (17, 23 kDa), calcium and chloride in photosynthetic oxygen evolution. *FEBS Letters* **192**, 1–3.
- **Ghanotakis DF, Babcock GT and Yocum CF** (1985*b*) Structure of the oxygen-evolving complex of photosystem-II calcium and lanthanum compete for sites on the oxidizing side of photosystem-II which control the binding of water-soluble polypeptides and regulate the activity of the manganese complex. *Biochimica et Biophysica Acta* **809**, 173–180.
- Goodin DB, Yachandra VK, Britt RD and Sauer K (1984) The state of manganese in the photosynthetic apparatus 3. Light-induced changes in X-ray absorption (K-edge) energies of manganese in photosynthetic membranes. *Biochimica et Biophysica Acta* 767, 209–216.
- Guo Y, Li H, He LL, Zhao DX, Gong LD and Yang ZZ (2017) The opencubane oxo-oxyl coupling mechanism dominates photosynthetic oxygen evolution: a comprehensive DFT investigation on O–O bond formation in the S-4 state. *Physical Chemistry Chemical Physics* **19**, 13909–13923.
- Haumann M and Junge W (1994) Extent and rate of proton release by photosynthetic water oxidation in thylakoids: electrostatic relaxation versus chemical production. *Biochemistry* **33**, 864–872.
- Haumann M, Bogershausen O, Cherepanov D, Ahlbrink R and Junge W (1997*a*) Photosynthetic oxygen evolution: H/D isotope effects and the coupling between electron and proton transfer during the redox reactions at the oxidizing side of photosystem II. *Photosynthesis Research* **51**, 193–208.
- Haumann M, Mulkidjanian AY and Junge W (1997*b*) The electrogenicities of electron and proton transfer at the oxidizing side of photosystem II. *Biochemistry* **36**, 9304–9315.
- Haumann M, Muller C, Liebisch P, Iuzzolino L, Dittmer J, Grabolle M, Neisius T, Meyer-Klaucke W and Dau H (2005) Structural and oxidation state changes of the photosystem II manganese complex in four transitions of the water oxidation cycle $(S-0\rightarrow S-1, S-1\rightarrow S-2, S-2\rightarrow S-3, and S-3, S-4\rightarrow S-0)$ characterized by X-ray absorption spectroscopy at 20 K and room temperature. *Biochemistry* **44**, 1894–1908.
- Haumann M, Grundmeier A, Zaharieva I and Dau H (2008) Photosynthetic water oxidation at elevated dioxygen partial pressure monitored by time-resolved X-ray absorption measurements. *Proceedings of the National Academy of Sciences of the United States of America* **105**, 17384–17389.
- Hays AM, Vassiliev IR, Golbeck JH and Debus RJ (1998) Role of D1-His190 in proton-coupled electron transfer reactions in photosystem II: a chemical complementation study. *Biochemistry* **37**, 11352–11365.
- Hellmich J, Bommer M, Burkhardt A, Ibrahim M, Kern J, Meents A, Muh F, Dobbek H and Zouni A (2014) Native-like photosystem II superstructure at 2.44 angstrom resolution through detergent extraction from the protein crystal. *Structure* **22**, 1607–1615.
- Hill R (1939) Oxygen produced by isolated chloroplasts. Proceedings of the Royal Society London 127, 192–210.
- Hillier W, Messinger J and Wydrzynski T (1998) Kinetic determination of the fast exchanging substrate water molecule in the S 3 state of photosystem II. *Biochemistry* 37, 16908–16914.
- Hoganson CW and Babcock GT (1997) A metalloradical mechanism for the generation of oxygen from water in photosynthesis. *Science* 277, 1953–1956.
- Hundelt M, Hays AM, Debus RJ and Junge W (1998) Oxygenic photosystem II: the mutation D1-D61N in Synechocystis sp. PCC 6803 retards S-state transitions without affecting electron transfer from Y Z to P 680+. *Biochemistry*, **37**, 14450–14456.

753

754

755

756

757

758

759

760

761

762

763

764

765

766

767

768

769

770

771

772

773

774

775

776

777

778

779

780

781

782

783

784

785

786

787

788

789

790

791

792

793

794

795

796

797

798

799

800

801

802

803

804

805

806

807

808

- Iuzzolino L, Dittmer J, D'RNER W, Meyer-Klaucke W and Dau H (1998) X-ray absorption spectroscopy on layered photosystem II membrane particles suggests manganese-centered oxidation of the oxygen-evolving complex for the S 0-S 1, S 1-S 2, and S 2-S 3 transitions of the water oxidizing complex. *Biochemistry* 37, 17112–17119.
- Iwata S and Barber J (2004) Structure of photosystem II and molecular architecture of the oxygen-evolving centre. *Current Opinion in Structural Biology* 14, 447–453.
- Jagendorf AT and Uribe E (1966) ATP formation caused by acid-base transition of spinach chloroplast. Proceedings of the National Academy of Sciences of the United States of America 55, 170–177.
- Joliot P and Joliot A (1968) A polarographic method for detection of oxygen production and reduction of Hill reagent by isolated chloroplasts. *Biochimica et Biophysica Acta* **153**, 625–634.
- Joliot P, Barbieri G and Chabaud R (1969) A new model of photochemical centers in system-2. *Photochemistry and Photobiology* **10**, 309.
- Jordan P, Fromme P, Witt HT, Klukas O, Saenger W and Krauss N (2001) Three-dimensional structure of cyanobacterial photosystem I at 2.5 angstrom resolution. *Nature* **411**, 909–917.
- Junge W (1970) Critical electric potential difference for photophosphorylation. Its relation to the chemiosmotic hypothesis and to the triggering requirements of the ATPase system. *European Journal of Biochemistry* 14, 582–592.
- Junge W (1977) Membrane potentials in photosynthesis. Annual Review of Plant Physiology and Plant Molecular Biology 28, 503–536.
- Junge W and Nelson N (2015) ATP synthase. Annual Review of Biochemistry 83, 631–657.
- Junge W and Rutherford AW (2007) Obituary: Horst Tobias Witt (1922–2007). Nature 448, 425.
- Junge W and Witt HT (1968) On the ion transport system of photosynthesis investigation on a molecular level. Zeitschrift für Naturforschung 23, 244–254.
- Junge W, Rumberg B and Schroeder H (1970) Necessity of an electric potential difference and its use for photophosphorylation in short flash groups. *European Journal of Biochemistry* 14, 575–581.
- Junge W, Haumann M, Ahlbrink R, Mulkidjanian A and Clausen J (2002) Electrostatics and proton transfer in photosynthetic water oxidation. *Philosophical Transactions of the Royal Society B: Biological Sciences* **357**, 1407–1417.
- Kamiya N and Shen JR (2003) Crystal structure of oxygen-evolving photosystem II from Thermosynechococcus vulcanus at 3.7-angstrom resolution. Proceedings of the National Academy of Sciences of the United States of America 100, 98–103.
- Kern J, Alonso-Mori R, Hellmich J, Tran R, Hattne J, Laksmono H, Glockner C, Echols N, Sierra RG, Sellberg J, Lassalle-Kaiser B, Gildea RJ, Glatzel P, Grosse-Kunstleve RW, Latimer MJ, Mcqueen TA, Difiore D, Fry AR, Messerschmidt M, Miahnahri A, Schafer DW, Seibert MM, Sokaras D, Weng TC, Zwart PH, White WE, Adams PD, Bogan MJ, Boutet S, Williams GJ, Messinger J, Sauter NK, Zouni A, Bergmann U, Yano J and Yachandra VK (2012) Room temperature femtosecond X-ray diffraction of photosystem II microcrystals. Proceedings of the National Academy of Sciences of the United States of America 109, 9721–9726.
- Kern J, Alonso-Mori R, Tran R, Hattne J, Gildea RJ, Echols N, Glockner C, Hellmich J, Laksmono H, Sierra RG, Lassalle-Kaiser B, Koroidov S, Lampe A, Han GY, Gul S, Difiore D, Milathianaki D, Fry AR, Miahnahri A, Schafer DW, Messerschmidt M, Seibert MM, Koglin JE, Sokaras D, Weng TC, Sellberg J, Latimer MJ, Grosse-Kunstleve RW, Zwart PH, White WE, Glatzel P, Adams PD, Bogan MJ, Williams GJ, Boutet S, Messinger J, Zouni A, Sauter NK, Yachandra VK, Bergmann U and Yano J (2013) Simultaneous femtosecond X-ray spectroscopy and diffraction of photosystem II at room temperature. *Science* 340, 491–495.
- Kern J, Tran R, Alonso-Mori R, Koroidov S, Echols N, Hattne J, Ibrahim M, Gul S, Laksmono H, Sierra RG, Gildea RJ, Han G, Hellmich J, Lassalle-Kaiser B, Chatterjee R, Brewster AS, Stan CA, Gloeckner C, Lampe A, Difiore D, Milathianaki D, Fry AR, Seibert MM, Koglin JE, Gallo E, Uhlig J, Sokaras D, Weng T-C,

Zwart PH, Skinner DE, Bogan MJ, Messerschmidt M, Glatzel P,
Williams GJ, Boutet S, Adams PD, Zouni A, Messinger J, Sauter NK,
Bergmann U, Yano J and Yachandra VK (2014) Taking snapshots of
photosynthetic water oxidation using femtosecond X-ray diffraction and
spectroscopy. Nature Communications 5.810811

- Kern J, Chatterjee R, Young ID, Fuller FD, Lassalle L, Ibrahim M, Gul S, Fransson T, Brewster AS, Alonso-Mori R, Hussein R, Zhang M, Douthit L, De Lichtenberg C, Cheah MH, Shevela D, Wersig J, Seuffert I, Sokaras D, Pastor E, Weninger C, Kroll T, Sierra RG, Aller P, Butryn A, Orville AM, Liang MN, Batyuk A, Koglin JE, Carbajo S, Boutet S, Moriarty NW, Holton JM, Dobbek H, Adams PD, Bergmann U, Sauter NK, Zouni A, Messinger J, Yano J and Yachandra VK (2018) Structures of the intermediates of Kok's photosynthetic water oxidation clock. *Nature* 563, 421.
- Kim CJ and Debus RJ (2017) Evidence from FTIR difference spectroscopy that a substrate H_2O molecule for O-2 formation in photosystem II is provided by the Ca Ion of the catalytic Mn_4CaO_5 cluster. *Biochemistry* 56, 2558–2570.
- Kirby JA, Goodin DB, Wydrzynski T, Robertson AS and Klein MP (1981a) State of manganese in the photosynthetic apparatus. 2. X-ray absorptionedge studies on manganese in photosynthetic membranes. *Journal of the American Chemical Society* **103**, 5537–5542.
- Kirby JA, Robertson AS, Smith JP, Thompson AC, Cooper SR and Klein MP (1981b) State of manganese in the photosynthetic apparatus. 1. Extended X-ray absorption fine-structure studies on chloroplasts and di-µ-oxo-bridged dimanganese model compounds. *Journal of the American Chemical Society* 103, 5529–5537.
- Klauss A, Krivanek R, Dau H and Haumann M (2009) Energetics and kinetics of photosynthetic water oxidation studied by photothermal beam deflection (PBD) experiments. *Photosynthesis Research*, 499–509.
- Klauss A, Haumann M and Dau H (2015) Seven steps of alternating electron and proton transfer in photosystem II water oxidation traced by timeresolved photothermal beam deflection at improved sensitivity. *Journal of Physical Chemistry B* **119**, 2677–2689.
- Kok B (1956) Photosynthesis in flashing light. *Biochimica et Biophysica Acta* 21, 245–258.
- Kok B and Gott W (1960) Activation spectra of 700-M-MU absorption change in photosynthesis. *Plant Physiology* **35**, 802–808.
- Kok B, Forbush B and Mcgloin M (1970) Cooperation of charges in photosynthetic O₂ evolution – I. A linear four-step mechanism. *Photochemistry* and Photobiology 11, 457–475.
- Kolling DR, Brown TS, Ananyev G and Dismukes GC (2009) Photosynthetic oxygen evolution is not reversed at high oxygen pressures: mechanistic consequences for the water-oxidizing complex 2. *Biochemistry* **48**, 1381–1389.
- Koroidov S, Shevela D, Shutova T, Samuelsson G and Messinger J (2014) Mobile hydrogen carbonate acts as proton acceptor in photosynthetic water oxidation. *Proceedings of the National Academy of Sciences of the United States of America* 111, 6299–6304.
- Krab K, Kempe H and Wikstrom M (2011) Explaining the enigmatic K(M) for oxygen in cytochrome c oxidase: a kinetic model. *Biochimica et Biophysica Acta* 1807, 348–358.
- Krewald V, Retegan M, Cox N, Messinger J, Lubitz W, Debeer S, Neese F and Pantazis DA (2015) Metal oxidation states in biological water splitting. *Chemical Science* 6, 1676–1695.
- Krewald V, Retegan M, Neese F, Lubitz W, Pantazis DA and Cox N (2016) Spin state as a marker for the structural evolution of nature's water splitting catalyst. *Inorganic Chemistry* 55, 488–501.
- Krishtalik LI (1989) Energetics of photosynthetic oxygen evolution. *Biofizika* 34, 883–886.
- Krishtalik LI (1990) Activation energy of photosynthetic oxygen evolution: an attempt at theoretical analysis. *Bioelectrochemistry and Bioenergetics* 23, 249–263.
- Krivanek R, Dau H and Haumann M (2008) Enthalpy changes during photosynthetic water oxidation tracked by time-resolved calorimetry using a photothermal beam deflection technique. *Biophysical Journal* 94, 1890–1903.
- **Kulik LV, Epel B, Lubitz W and Messinger J** (2005) Mn-55 pulse ENDOR at 34 GHz of the S-0 and S-2 states of the oxygen-evolving complex in photosystem II. *Journal of the American Chemical Society* **127**, 2392–2393.

825

826

827

828

829

830 831 832

833 834

835

836

837 838

> 839 840 841

842

843

844

845

846

847

848

849

850

851

852

853

854

855

856

857

858

859

860

861

862

863

864

865

866

867

868

869

870

Kump LR (2008) The rise of atmospheric oxygen. Nature 451, 277-278.

- Kupitz C, Basu S, Grotjohann I, Fromme R, Zatsepin NA, Rendek KN, Hunter MS, Shoeman RL, White TA, Wang DJ, James D, Yang JH, Cobb DE, Reeder B, Sierra RG, Liu HG, Barty A, Aquila AL, Deponte D, Kirian RA, Bari S, Bergkamp JJ, Beyerlein KR, Bogan MJ, Caleman C, Chao TC, Conrad CE, Davis KM, Fleckenstein H, Galli L, Hau-Riege SP, Kassemeyer S, Laksmono H, Liang MN, Lomb L, Marchesini S, Martin AV, Messerschmidt M, Milathianaki D, Nass K, Ros A, Roy-Chowdhury S, Schmidt K, Seibert M, Steinbrener J, Stellato F, Yan LF, Yoon C, Moore TA, Moore AL, Pushkar Y, Williams GJ, Boutet S, Doak RB, Weierstall U, Frank M, Chapman HN, Spence JCH and Fromme P (2014) Serial time-resolved crystallography of photosystem II using a femtosecond X-ray laser. Nature 513, 261.
- Kusunoki M (2007) Mono-manganese mechanism of the photosystem II water splitting reaction by a unique Mn4Ca cluster. *Biochimica et Biophysica Acta-Bioenergetics* 1767, 484–492.
- Kusunoki M (2011) S-1-state Mn4Ca complex of photosystem II exists in equilibrium between the two most-stable isomeric substates: XRD and EXAFS evidence. *Journal of Photochemistry and Photobiology B-Biology* 104, 100–110.
- Latimer MJ, Derose VJ, Mukerji I, Yachandra VK, Sauer K and Klein MP (1995) Evidence for the proximity of calcium to the manganese cluster of photosystem II: determination by X-ray absorption spectroscopy. *Biochemistry* 34, 10898–10909.
- Lavergne J and Junge W (1993) Proton release during the redox cycle of the water oxidase. *Photosynthesis Research* 38, 279–296.
- Li XC, Sproviero EM, Ryde U, Batista VS and Chen GJ (2013) Theoretical EXAFS studies of a model of the oxygen-evolving complex of photosystem II obtained with the quantum cluster approach. *International Journal of Quantum Chemistry* **113**, 474–478.
- Loll B, Kern J, Saenger W, Zouni A and Biesiadka J (2005) Towards complete cofactor arrangement in the 3.0 angstrom resolution structure of photosystem II. *Nature* **438**, 1040–1044.
- Mcevoy JP, Gascon JA, Batista VS and Brudvig GW (2005) The mechanism of photosynthetic water splitting. *Photochemical & Photobiological Sciences* **4**, 940–949.
- Menke W (1962) Structure and chemistry of plastids. *Annual Review of Plant Physiology* **13**, 27–44.
- Messinger J (2000) Towards understanding the chemistry of photosynthetic oxygen evolution: dynamic structural changes, redox states and substrate water binding of the Mn cluster in photosystem II. *Biochimica et Biophysica Acta-Bioenergetics* 1459, 481–488.
- Messinger J (2004) Evaluation of different mechanistic proposals for water oxidation in photosynthesis on the basis of Mn₄O_xCa structures for the catalytic site and spectroscopic data. *Physical Chemistry Chemical Physics* 6, 4764–4771.
- Messinger J, Badger M and Wydrzynski T (1995) Detection of one slowly exchanging substrate water molecule in the S 3 state of photosystem II. Proceedings of the National Academy of Sciences of the United States of America 92, 3209–3213.
- Messinger J, Robblee JH, Bergmann U, Fernandez C, Glatzel P, Visser H, Cinco RM, Mcfarlane KL, Bellacchio E, Pizarro SA, Cramer SP, Sauer K, Klein MP and Yachandra VK (2001) Absence of Mn-centered oxidation in the S-2→S-3 transition: implications for the mechanism of photosynthetic water oxidation. *Journal of the American Chemical Society* 123, 7804–7820.
- Meyer B, Schlodder E, Dekker JP and Witt HT (1989) O₂ evolution and Chl a II + (P-680+) nanosecond reduction kinetics in single flashes as a function of pH. *Biochimica et Biophysica Acta* **974**, 36–43.
- Mitchell P (1961) Coupling of photophosphorylation to electron and hydrogen transfer by a chemiosmotic type of mechanism. *Nature* **191**, 144–148.
- Mitchell P (1966) Chemiosmotic coupling in oxidative and photosynthetic phosphorylation. *Physiological Reviews* **41**, 445–502.
- Narzi D, Bovi D and Guidoni L (2014) Pathway for Mn-cluster oxidation by tyrosine-Z in the S-2 state of photosystem II. Proceedings of the National Academy of Sciences of the United States of America 111, 8723–8728.
- Nilsson H, Krupnik T, Kargul J and Messinger J (2014a) Substrate water exchange in photosystem II core complexes of the extremophilic red alga

Cyanidioschyzon merolae. Biochimica et Biophysica Acta-Bioenergetics **1837**, 1257–1262.

- Nilsson H, Rappaport F, Boussac A and Messinger J (2014b) Substrate-water exchange in photosystem II is arrested before dioxygen formation. *Nature Communications* 5:4305, 1–7.
- Nilsson H, Cournac L, Rappaport F, Messinger J and Lavergne J (2016) Estimation of the driving force for dioxygen formation in photosynthesis. *Biochimica et Biophysica Acta-Bioenergetics* **1857**, 23–33.
- Nitschke W and Rutherford AW (1991) Photosynthetic reaction centers: variations on a common structural theme? *Trends in Biochemical Sciences* 16, 241–245.
- **Orsi F, Muratori M, Rocco M, Colombo E and Rizzoni G** (2016) A multidimensional well-to-wheels analysis of passenger vehicles in different regions: primary energy consumption, CO₂ emissions, and economic cost. *Applied Energy* **169**, 197–209.
- Ort DR, Merchant SS, Alric J, Barkan A, Blankenship RE, Bock R, Croce R, Hanson MR, Hibberd JM, Long SP, Moore TA, Moroney J, Niyogi KK, Parry MAJ, Peralta-Yahya PP, Prince RC, Redding KE, Spalding MH, Van Wijk KJ, Vermaas WFJ, Von Caemmerer S, Weber APM, Yeates TO, Yuan JS and Zhu XG (2015) Redesigning photosynthesis to sustainably meet global food and bioenergy demand. Proceedings of the National Academy of Sciences of the United States of America 112, 8529– 8536.
- Pecoraro VL, Baldwin MJ, Caudle MT, Hsieh WY and Law NA (1998) A proposal for water oxidation in photosystem II. *Pure and Applied Chemistry* **70**, 925–929.
- **Peloquin JM and Britt RD** (2001) EPR/ENDOR characterization of the physical and electronic structure of the OEC Mn cluster. *Biochimica et Biophysica Acta* **1503**, 96–111.
- Petrie S, Gatt P, Stranger R and Pace RJ (2012) Modelling the metal atom positions of the photosystem II water oxidising complex: a density functional theory appraisal of the 1.9 angstrom resolution crystal structure. *Physical Chemistry Chemical Physics* 14, 11333–11343.
- Planavsky NJ, Asael D, Hofmann A, Reinhard CT, Lalonde SV, Knudsen A, Wang XL, Ossa FO, Pecoits E, Smith AJB, Beukes NJ, Bekker A, Johnson TM, Konhauser KO, Lyons TW and Rouxel OJ (2014) Evidence for oxygenic photosynthesis half a billion years before the great oxidation event. *Nature Geoscience* 7, 283–286.
- **Priestley J** (1772) Observations of different kinds of air. *Philosophical Transactions of the Royal Society of London* **62**, 147–264.
- Rapatskiy L, Cox N, Savitsky A, Ames WM, Sander J, Nowaczyk MM, Rogner M, Boussac A, Neese F, Messinger J and Lubitz W (2012) Detection of the water-binding sites of the oxygen-evolving complex of photosystem II using W-band 17O electron-electron double resonancedetected NMR spectroscopy. *Journal of the American Chemical Society* 134, 16619–16634.
- **Raven JA and Larkum AWD** (2007) Are there ecological implications for the proposed energetic restrictions on photosynthetic oxygen evolution at high oxygen concentrations? *Photosynthesis Research* **94**, 31–42.
- **Renger G** (1977) A model for the molecular mechanism of photosynthetic oxygen evolution. *FEBS Letters* **81**, 223–228.
- **Renger G** (1987) Mechanistic aspects of photosynthetic water cleavage. *Photosynthetica* **21**, 203–224.
- Retegan M, Cox N, Lubitz W, Neese F and Pantazis DA (2014) The first tyrosyl radical intermediate formed in the S-2-S-3 transition of photosystem II. *Physical Chemistry Chemical Physics* **16**, 11901–11910.
- Retegan M, Krewald V, Mamedov F, Neese F, Lubitz W, Cox N and Pantazis DA (2016) A five-coordinate Mn(IV) intermediate in biological water oxidation: spectroscopic signature and a pivot mechanism for water binding. *Chemical Science* 7, 72–84.
- Rhee KH, Morriss EP, Barber J and Kuhlbrandt W (1998) Threedimensional structure of the plant photosystem II reaction centre at 8 angstrom resolution. *Nature* **396**, 283–286.
- **Rieger B, Junge W and Busch KB** (2014) Lateral pH gradient between OXPHOS complex IV and F(0)F(1) ATP-synthase in folded mitochondrial membranes. *Nature Communications* **5**, 3103.
- Robblee JH, Messinger J, Cinco RM, Mcfarlane KL, Fernandez C, Pizarro SA, Sauer K and Yachandra VK (2002) The Mn cluster in the 933

881

882

872

873

874

875

876

887

888

889

890

891

892

893

894

895

896

897

898

899

900

901

902

903

904

905

906

907

908

909

910

911

912

913

914

915

916

917

918

919

920

921

922

923

924

925

926

927

928

929

930

934

935

936

937

938

939

940

941

942

943

944

945

946

947

948

949

950

951

952

953

954

955

956

957

958

959

960

961

962

963

964

965

966

967

968

969

970

971

972

973

974

975

976

977

978

979

980

981

982

983

984

985

986

987

988

989

990

991

992

993

994

995

S-0 state of the oxygen-evolving complex of photosystem II studied by EXAFS spectroscopy: are there three di-mu-oxo-bridged Mn-2 moieties in the tetranuclear Mn complex? *Journal of the American Chemical Society* **124**, 7459–7471.

- Roelofs TA, Liang W, Latimer MJ, Cinco RM, Rompel A, Andrews JC, Sauer K, Yachandra VK and Klein M (1996) Oxidation states of the manganese cluster during the flash-induced S-state cycle of the photosynthetic oxygen-evolving complex. Proceedings of the National Academy of Sciences of the United States of America 93, 3335–3340.
- Rögner M, Dekker JP, Boekema EJ and Witt HT (1987) Size, shape and mass of the oxygen-evolving photosystem II complex from the thermophilic cyanobacterium Synechococcus sp. FEBS Letters 219, 207–211.
- **Ruben SR, Kamen M and Hyde JL** (1941) Heavy oxygen (O¹⁸) as a tracer in the study of photosynthesis. *Journal of the American Chemical Society* **63**, 877–879.

Rutherford AW, Osyczka A and Rappaport F (2012) Back-reactions, shortcircuits, leaks and other energy wasteful reactions in biological electron transfer: redox tuning to survive life in O(2). FEBS Letters 586, 603–616.

- Saito T, Yamanaka S, Kanda K, Isobe H, Takano Y, Shigeta Y, Umena Y, Kawakami K, Shen JR, Kamiya N, Okumura M, Shoji M, Yoshioka Y and Yamaguchi K (2012) Possible mechanisms of water splitting reaction based on proton and electron release pathways revealed for CaMn₄O₅ cluster of PSII refined to 1.9 angstrom X-ray resolution. *International Journal of Quantum Chemistry* 112, 253–276.
- Sakamoto H, Shimizu T, Nagao R and Noguchi T (2017) Monitoring the reaction process during the S-2→ S-3 transition in photosynthetic water oxidation using time-resolved infrared spectroscopy. *Journal of the American Chemical Society* **139**, 2022–2029.
- Sauer K (1980) A role for manganese in oxygen evolution in photosynthesis. Accounts of Chemical Research 13, 249–256.
- Schatz GH, Brock H and Holzwarth AR (1987) Picosecond kinetics of fluorescence and absorbency changes in photosystem-ii particles excited at low photon density. *Proceedings of the National Academy of Sciences of the United States of America* 84, 8414–8418.

Schatz GH, Brock H and Holzwarth AR (1988) Kinetic and energetic model for the primary processes in photosystem-ii. *Biophysical Journal* 54, 397–405.

- Schiller H, Dittmer J, Iuzzolino L, Dorner W, Meyer-Klaucke W, Sole VA, Nolting HF and Dau H (1998) Structure and orientation of the oxygenevolving manganese complex of green algae and higher plants investigated by X-ray absorption linear dichroism spectroscopy on oriented photosystem II membrane particles. *Biochemistry* 37, 7340–7350.
- Schliephake W, Junge W and Witt HT (1968) Correlation between field formation, proton translocation, and the light reactions in photosynthesis. *Zeitschrift für Naturforschung* 23, 1571–1578.
- Schlodder E, Brettel K, Schatz GH and Witt HT (1984) Analysis of the Chl- a II+ reduction kinetics with nanosecond time resolution in oxygen-evolving photosystem II particles from Synechococcus at 680 and 824 nm. *Biochimica et Biophysica Acta* 765, 178–185.
- Schönknecht G, Althoff G and Junge W (1990) The electric unit size of thylakoid membranes. FEBS Letters 277, 65–68.
- Schuth N, Zaharieva I, Chernev P, Berggren G, Anderlund M, Styring S, Dau H and Haumann M (2018) K alpha X-ray emission spectroscopy on the photosynthetic oxygen-evolving complex supports manganese oxidation and water binding in the S-3 state. *Inorganic Chemistry* 57, 10424–10430.
- Shen JR and Kamiya N (2000) Crystallization and the crystal properties of the oxygen-evolving photosystem II from Synechococcus vulcanus. *Biochemistry* 39, 14739–14744.
- Shevela D, Beckmann K, Clausen J, Junge W and Messinger J (2011) Membrane-inlet mass spectrometry reveals a high driving force for oxygen production by photosystem II. Proceedings of the National Academy of Sciences of the United States of America 108, 3602–3607.
- Shevela D, Eaton-Rye JJ, Shen JR and Govindjee G (2012) Photosystem II and the unique role of bicarbonate: a historical perspective. *Biochimica et Biophysica Acta* 1817, 1134–1151.
- Siegbahn PE (2008a) Mechanism and energy diagram for O–O bond formation in the oxygen-evolving complex in photosystem II. *Philosophical Transactions of the Royal Society B: Biological Sciences* 363, 1221–1228.

- Siegbahn PE (2008b) A structure-consistent mechanism for dioxygen formation in photosystem II. Chemistry 14, 8290–8302.
- Siegbahn PE (2009) Structures and energetics for O₂ formation in photosystem II. Accounts of Chemical Research 42, 1871–1880.
- Siegbahn PEM (2000) Theoretical models for the oxygen radical mechanism of water oxidation and of the water oxidizing complex of photosystem II. *Inorganic Chemistry* 39, 2923–2935.
- Siegbahn PEM (2006) O–O bond formation in the S-4 state of the oxygenevolving complex in photosystem II. *Chemistry-A European Journal* 12, 9217–9227.
- Siegbahn PEM (2013*a*) Substrate water exchange for the oxygen evolving complex in PSII in the S-1, S-2, and S-3 states. *Journal of the American Chemical Society* 135, 9442–9449.
- Siegbahn PEM (2013*b*) Water oxidation mechanism in photosystem II, including oxidations, proton release pathways, O–O bond formation and O-2 release. *Biochimica et Biophysica Acta-Bioenergetics* **1827**, 1003–1019.
- **Siegbahn PEM** (2017) Nucleophilic water attack is not a possible mechanism for O-O bond formation in photosystem II. *Proceedings of the National Academy of Sciences of the United States of America* **114**, 4966–4968.
- Siegbahn PEM and Blomberg MRA (2014) Energy diagrams for water oxidation in photosystem II using different density functionals. *Journal of Chemical Theory and Computation* **10**, 268–272.
- Siegbahn PEM and Crabtree RH (1999) Manganese oxyl radical intermediates and O–O bond formation in photosynthetic oxygen evolution and a proposed role for the calcium cofactor in photosystem II. *Journal of the American Chemical Society* 121, 117–127.
- Sjoholm J, Bergstrand J, Nilsson T, Sachl R, Von Ballmoos C, Widengren J and Brzezinski P (2017) The lateral distance between a proton pump and ATP synthase determines the ATP-synthesis rate. *Scientific Reports* 7:2926, 1–12.
- South PF, Cavanagh AP, Liu HW and Ort DR (2018) Improving crop productivity by bypassing photorespiration: a synthetic biology approach. In Vitro Cellular & Developmental Biology-Animal 54, S18–S19.
- **Spector MB and Winget GD** (1979) Chloroplast membrane-protein required for the photosynthetic oxidation of water. *Ohio Journal of Science* **79**, 21–21.
- Sproviero EM, Gascon JA, Mcevoy JP, Brudvig GW and Batista VS (2006) QM/MM models of the O-2-evolving complex of photosystem II. *Journal of Chemical Theory and Computation* 2, 1119–1134.
- Sproviero EM, Gascon JA, Mcevoy JP, Brudvig GW and Batista VS (2008) Quantum mechanics/molecular mechanics study of the catalytic cycle of water splitting in photosystem II. *Journal of the American Chemical Society* **130**, 3428–3442.
- Suga M, Akita F, Hirata K, Ueno G, Murakami H, Nakajima Y, Shimizu T, Yamashita K, Yamamoto M, Ago H and Shen J-R (2015) Native structure of photosystem II at 1.95 angstrom resolution viewed by femtosecond X-ray pulses. *Nature* 517, 99–U265.
- Suga M, Akita F, Sugahara M, Kubo M, Nakajima Y, Nakane T, Yamashita K, Umena Y, Nakabayashi M, Yamane T, Nakano T, Suzuki M, Masuda T, Inoue S, Kimura T, Nomura T, Yonekura S, Yu LJ, Sakamoto T, Motomura T, Chen JH, Kato Y, Noguchi T, Tono K, Joti Y, Kameshima T, Hatsui T, Nango E, Tanaka R, Naitow H, Matsuura Y, Yamashita A, Yamamoto M, Nureki O, Yabashi M, Ishikawa T, Iwata S and Shen JR (2017) Light-induced structural changes and the site of O=O bond formation in PSII caught by XFEL. Nature 543, 131.
- Suzuki H, Sugiura M and Noguchi T (2008) Monitoring water reactions during the S-state cycle of the photosynthetic water-oxidizing center: detection of the DOD bending vibrations by means of Fourier transform infrared spectroscopy. *Biochemistry* 47, 11024–11030.
- Tanaka A, Fukushima Y and Kamiya N (2017) Two different structures of the oxygen-evolving complex in the same polypeptide frameworks of photosystem II. *Journal of the American Chemical Society* 139, 1718–1721.
- Tommos C and Babcock GT (1998) Oxygen production in nature: a light-driven metalloradical enzyme process. *Accounts of Chemical Research* **31**, 18–25.
- Torchio MF and Santarelli MG (2010) Energy, environmental and economic comparison of different powertrain/fuel options using well-to-wheels assessment, energy and external costs European market analysis. *Energy* **35**, 4156–4171.
- Tran R, Kern J, Hattne J, Koroidov S, Hellmich J, Alonso-Mori R, Sauter NK, Bergmann U, Messinger J, Zouni A, Yano J and

Yachandra VK (2014) The Mn₄Ca photosynthetic water-oxidation catalyst studied by simultaneous X-ray spectroscopy and crystallography using an X-ray free-electron laser. Philosophical Transactions of the Royal Society B-Biological Sciences 369:20130324, 1-5.

- Umena Y, Kawakami K, Shen J-R and Kamiya N (2011) Crystal structure of oxygen-evolving photosystem II at a resolution of 1.9 angstrom. Nature 473, 55-U65.
- Wang M, Han J, Dunn JB, Cai H and Elgowainy A (2012) Well-to-wheels energy use and greenhouse gas emissions of ethanol from corn, sugarcane and cellulosic biomass for US use. Environmental Research Letters 7:045905, 1-13
- Warburg ONE (1922). Über den Energieumsatz bei der Kohlensäureassimilation. Zeitschrift für Physikalische Chemie 102, 235-266.
- Warburg O, Krippahl G and Jetschma C (1965) Widerlegung der Photolyse des Wassers und Beweis der Photolyse der Kohlensäure nach Versuchen mit lebender Chlorella und den Hill-Reagentien Nitrat und K3Fe(Cn)6. Zeitschrift für Naturforschung B 20, 993–996.
- Wieghardt K (1989) The active-sites in manganese-containing metalloproteins and inorganic model complexes. Angewandte Chemie-International Edition in English 28, 1153-1172.
- Witt HT and Moraw R (1959) Untersuchungen über die Primärvorgänge bei der Photosynthese. I. Mitteilung. Zeitschrift für Physikalische Chemie 20, 253 - 282.
- Witt HT, Mueller A and Rumberg B (1961a) Experimental evidence for the mechanism of photosynthesis. Nature 191, 194-195.
- Witt HT, Mueller A and Rumberg B (1961b) Oxidized cytochrome and chlorophyll C2-plus in photosynthesis. Nature 192, 967-969.
- Witt I, Witt HT, Difiore D, Rogner M, Hinrichs W, Saenger W, Granzin J, Betzel C and Dauter Z (1988) X-ray characterization of single-crystals of the reaction center-I of water splitting photosynthesis. Berichte Der Bunsen-Gesellschaft-Physical Chemistry Chemical Physics 92, 1503-1506.
- Wydrzynski T and Sauer K (1980) Periodic changes in the oxidation-state of manganese in photosynthetic oxygen evolution upon illumination with flashes. Biochimica et Biophysica Acta 589, 56-70.
- Yachandra VK, Guiles RD, Mcdermott AE, Cole JL, Britt RD, Dexheimer SL, Sauer K and Klein MP (1987a) Comparison of the structure of the manganese complex in the S1 and S2 states of the photosynthetic O-2-evolving complex: an X-ray absorption spectroscopy study. Biochemistry 26, 5974-5981.
- Yachandra VK, Guiles RD, Mcdermott AE, Cole JL, Britt RD, Dexheimer SL, Sauer K and Klein MP (1987b) The state of manganese in the photosynthetic apparatus. 7. Comparison of the structure of the manganese complex in the s-1 and s-2 states of the photosynthetic o-2evolving complex - an X-ray absorption-spectroscopy study. Biochemistry 26, 5974-5981.
- Yachandra VK, Derose VJ, Latimer MJ, Mukerji I, Sauer K and Klein MP (1993) Where plants make oxygen: a structural model for the photosynthetic oxygen-evolving manganese cluster. Science 260, 675-679.
- Yachandra VK, Sauer K and Klein MP (1996) Manganese cluster in photosynthesis: where plants oxidize water to dioxygen. Chemical Reviews 96, 2927-2950.
- Yamanaka S, Isobe H, Kanda K, Saito T, Umena Y, Kawakami K, Shen JR, Kamiya N, Okumura M, Nakamura H and Yamaguchi K (2011) Possible mechanisms for the O-O bond formation in oxygen evolution reaction at the CaMn₄O₅(H₂O)(4) cluster of PSII refined to 1.9 angstrom X-ray resolution. Chemical Physics Letters 511, 138–145.

- Yano J, Kern J, Irrgang KD, Latimer MJ, Bergmann U, Glatzel P, Pushkar Y, Biesiadka J, Loll B, Sauer K, Messinger J, Zouni A and Yachandra VK (2005) X-ray damage to the Mn4Ca complex in single crystals of photosystem II: a case study for metalloprotein crystallography. Proceedings of the National Academy of Sciences of the United States of 1000 America 102, 12047-12052. 1001
- Yano J, Kern J, Sauer K, Latimer MJ, Pushkar Y, Biesiadka J, Loll B, Saenger W, Messinger J, Zouni A and Yachandra VK (2006) Where water is oxidized to dioxygen: structure of the photosynthetic Mn4Ca cluster. Science 314, 821-825.
- Yocum CF (1991) Calcium activation of photosynthetic water oxidation. Biochimica et Biophysica Acta 1059, 1-15.
- Young ID, Ibrahim M, Chatterjee R, Gul S, Fuller FD, Koroidov S, 1007 Brewster AS, Tran R, Alonso-Mori R, Kroll T, Michels-Clark T, 1008 Laksmono H, Sierra RG, Stan CA, Hussein R, Zhang M, Douthit L, 1009 Kubin M, De Lichtenberg C, Pham LV, Nilsson H, Cheah MH, 1010 Shevela D, Saracini C, Bean MA, Seuffert I, Sokaras D, Weng TC, 1011 Pastor E, Weninger C, Fransson T, Lassalle L, Brauer P, Aller P, 1012 Docker PT, Andi B, Orville AM, Glownia JM, Nelson S, Sikorski M, 1013 Zhu DL, Hunter MS, Lane TJ, Aquila A, Koglin JE, Robinson J, Liang MN, Boutet S, Lyubimov AY, Uervirojnangkoorn M, 1014 Moriarty NW, Liebschner D, Afonine PV, Waterman DG, Evans G, 1015 Wernet P, Dobbek H, Weis WI, Brunger AT, Zwart PH, Adams PD, 1016 Zouni A, Messinger J, Bergmann U, Sauter NK, Kern J, Yachandra VK 1017 and Yano J (2016) Structure of photosystem II and substrate binding at 1018 room temperature. Nature 540, 453. 1019
- Zaharieva I, Chernev P, Berggren G, Anderlund M, Styring S, Dau H and 1020 Haumann M (2016a) Room-temperature energy-sampling K beta X-ray 1021 emission spectroscopy of the Mn4Ca complex of photosynthesis reveals 1022 three manganese-centered oxidation steps and suggests a coordination 1023 change prior to O-2 formation. Biochemistry 55, 4197-4211. 1024
- Zaharieva I, Dau H and Haumann M (2016b) Sequential and coupled proton and electron transfer events in the S-2 \rightarrow S-3 transition of photosynthetic water oxidation revealed by time-resolved X-ray absorption spectroscopy. Biochemistry 55, 6996-7004.
- Zhang CX, Chen CH, Dong HX, Shen JR, Dau H and Zhao JQ (2015) A synthetic Mn4Ca-cluster mimicking the oxygen-evolving center of photosynthesis. Science 348, 690-693.
- Zhang M, Bommer M, Chatterjee R, Hussein R, Yano J, Dau H, Kern J, Dobbek H and Zouni A (2017) Structural insights into the light-driven auto-assembly process of the water oxidizing Mn₄CaO₅-cluster in photosystem II. Elife 6:e26933, 1-20.
- Zheng M and Dismukes GC (1996) Orbital configuration of the valence electrons, ligand field symmetry, and manganese oxidation states of the photosynthetic water oxidizing complex: analysis of the S2 state multiline EPR signals. Inorganic Chemistry 35, 3307-3319.
- Zhu XG, Long SP and Ort DR (2008) What is the maximum efficiency with 1038 which photosynthesis can convert solar energy into biomass? Current 1039 Opinion in Biotechnology 19, 153-159.
- 1040 Zouni A, Jordan R, Schlodder E, Fromme P and Witt HT (2000) First 1041 photosystem II crystals capable of water oxidation. Biochimica et 1042 Biophysica Acta 1457, 103-105.
- Zouni A, Witt HT, Kern J, Fromme P, Krauss N, Saenger W and Orth P (2001) Crystal structure of photosystem II from Synechococcus elongatus at 3.8 resolution. Nature 409, 739-743.

996

997

998

999

1002

1003

1004

1005

1006

1025

1026

1027

1028

1029

1030

1031

1032

1033

1034

1035

1036

1037

1043

1044